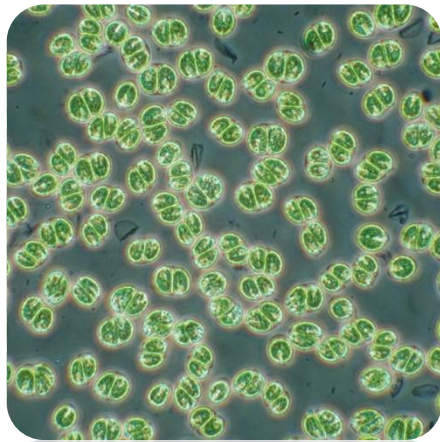


PRODUCTION OF STABLE MICROALGAE-ENRICHED FOAM

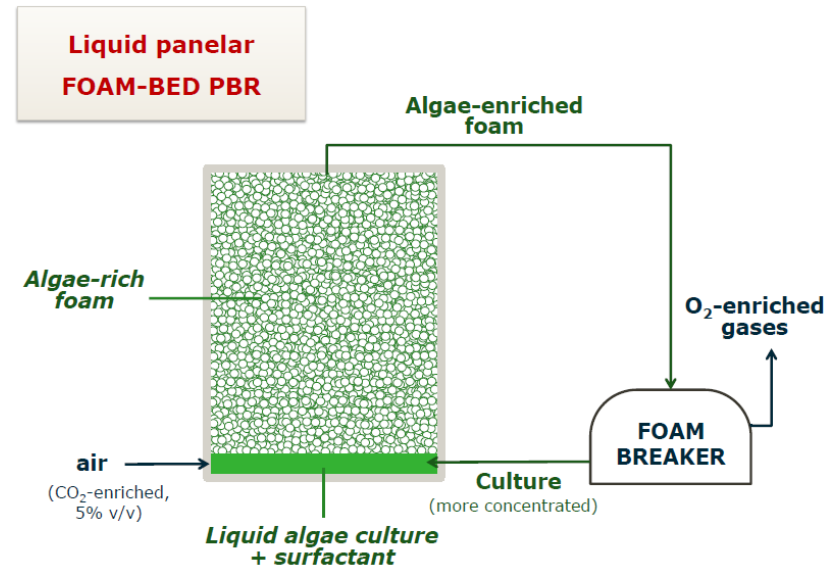
M. Vázquez, V. Si Atat, I. Garbayo, M. Cuaresma, C. Vílchez
University of Huelva



AIM

Main objective

- Development of a novel **Foam-Bed Photobioreactor** for cost reduction of microalgal biomass production



Specific aims (foam formation)

- Selection of suitable **microalgae strains**
- Determine **optimal culture conditions** to produce stable algae-rich foam

JUSTIFICATION

Why to cultivate algae in foam?

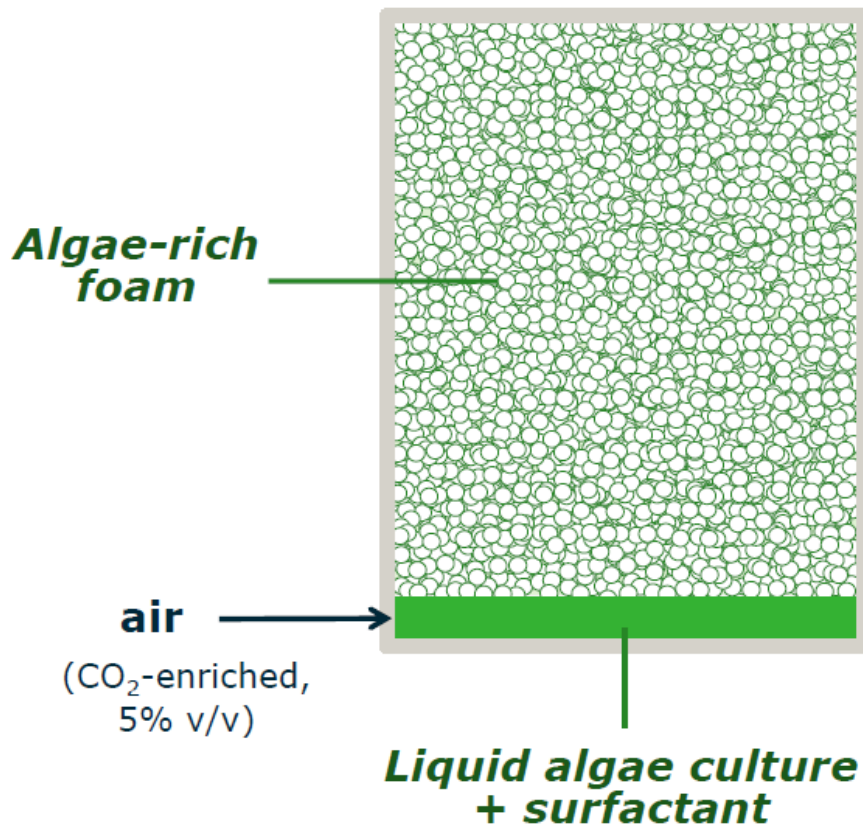
- A foam-bed PBR is an attempt for **reducing cost of algae production** processes.
- ✓ *High biomass productivities*
- ✓ *Improved mass transfer*
- ✓ *Improved light use efficiency*
- ✓ *Dramatic reduction of water usage*
- ✓ *Harvesting costs reduction*
- ✓ *Avoid dewatering processes like centrifugation, filtration or flocculation*



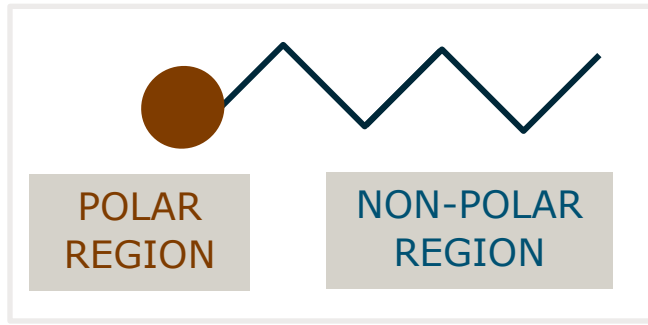
BACKGROUND

Foam flotation as an algal production system

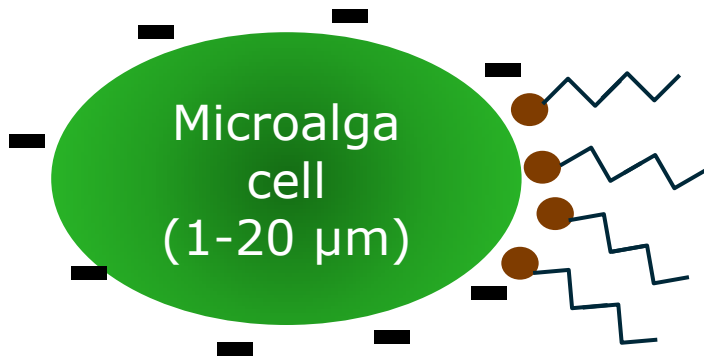
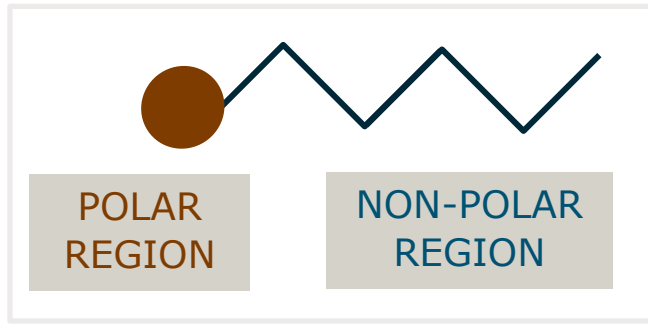
Until now, foam flotation has been used as a harvesting method (but not for cultivation!):



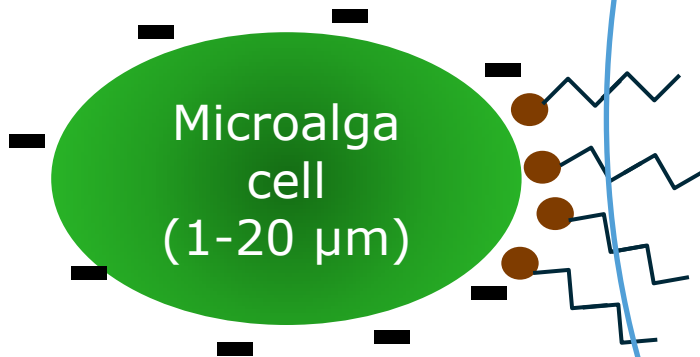
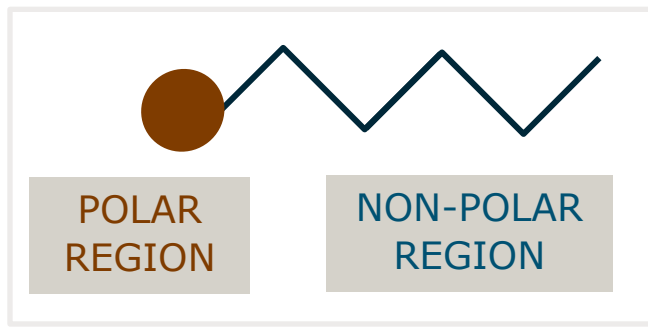
BACKGROUND



BACKGROUND

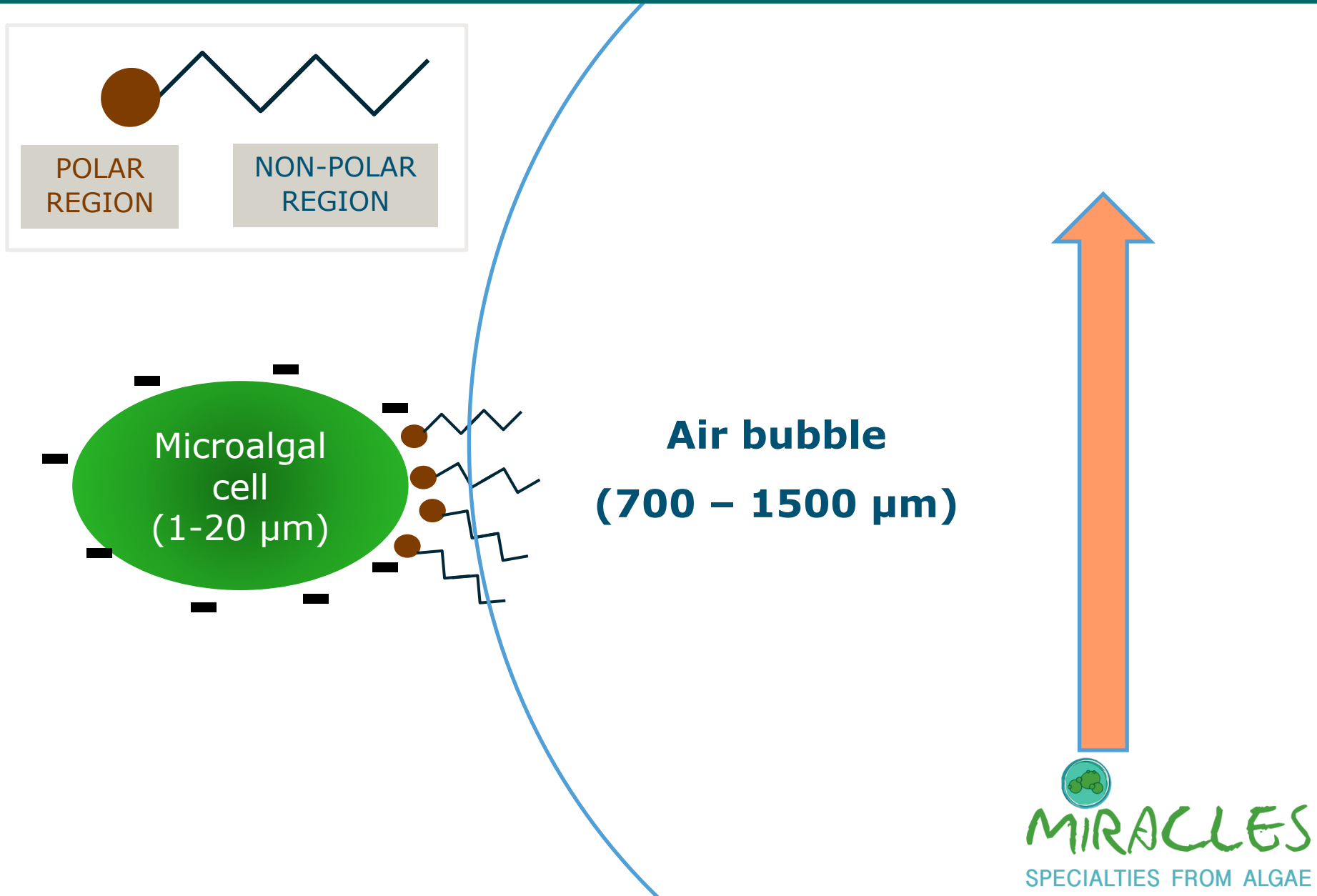


BACKGROUND

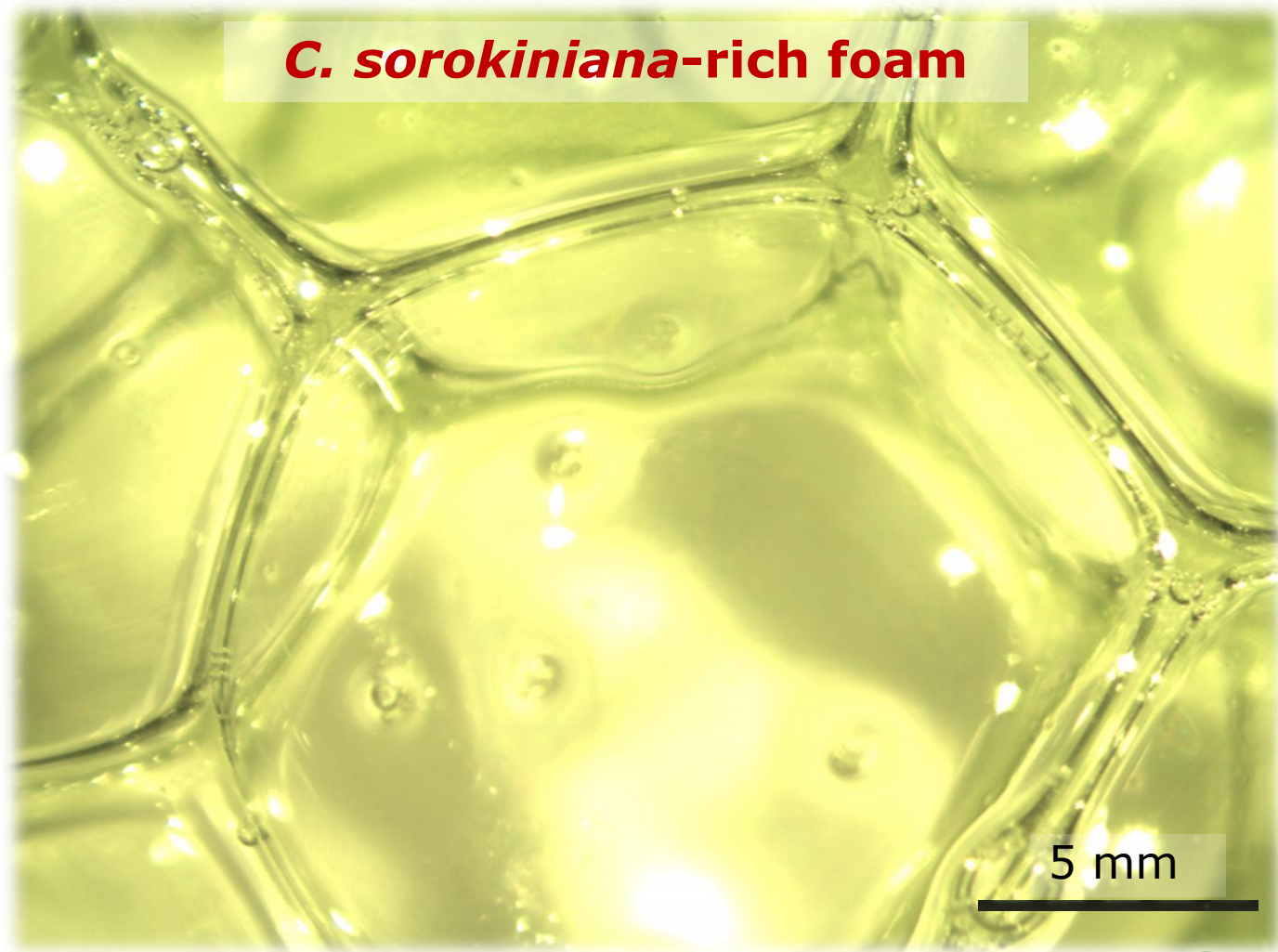


Air bubble
(700 – 1500 μm)

BACKGROUND



BACKGROUND

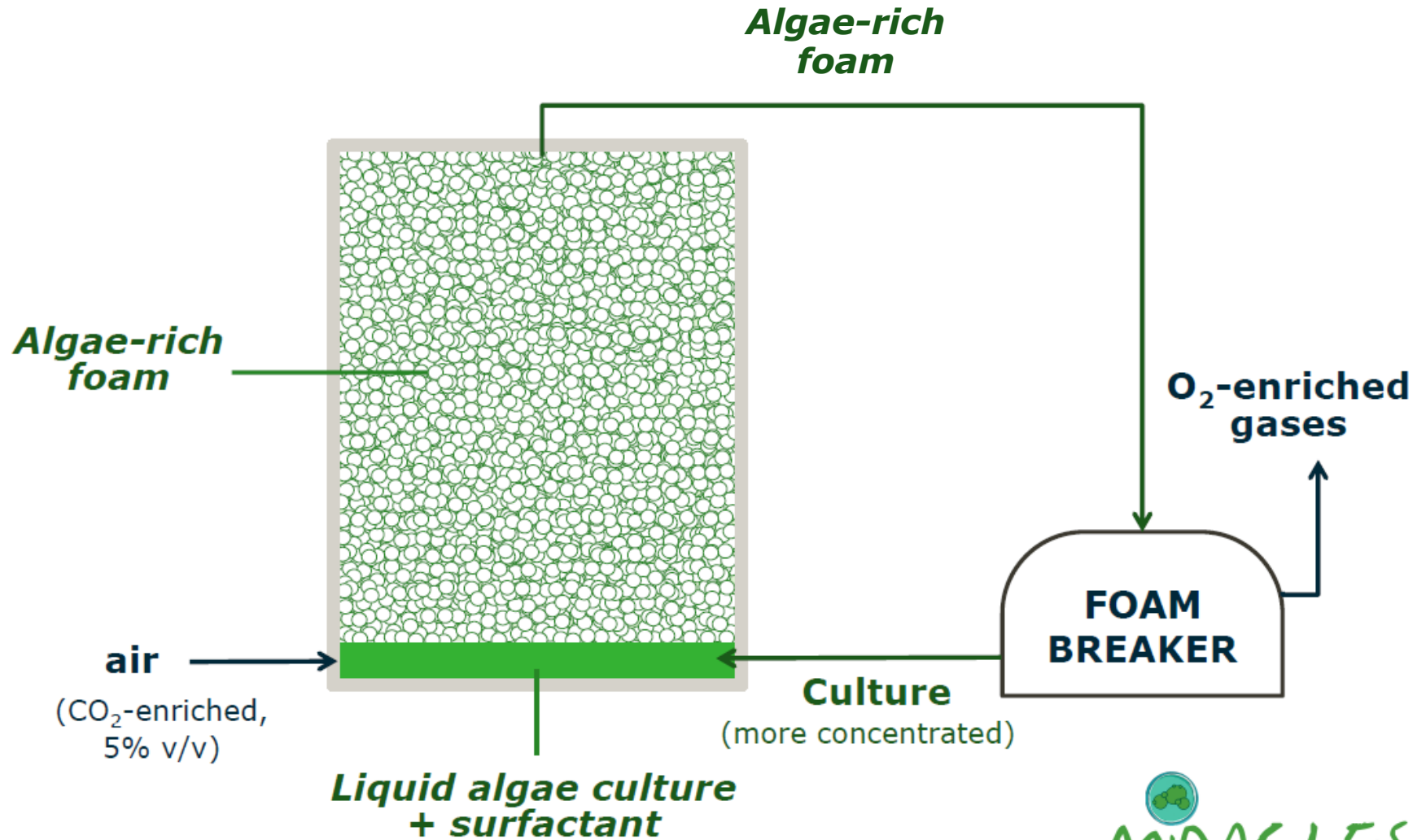


***C. sorokiniana*-rich foam**

5 mm

Magnifying glass
3x10

BACKGROUND



BACKGROUND

What are the desirable conditions for producing algae-rich foam?

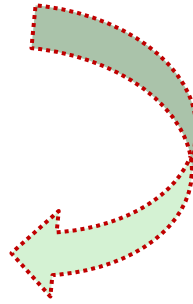
- ✓ Suitable choice of surfactant + microalga strain
- ✓ Microalgae hold-up → Algae-enriched foam
- ✓ Microalgae viability
- ✓ Stable foam
- ✓ Net foam accumulation



BACKGROUND

What are the desirable conditions for producing algae-rich foam?

- ✓ Suitable choice of surfactant + microalga strain
- ✓ Microalgae hold-up → Algae-enriched foam
- ✓ Microalgae viability
- ✓ Stable foam
- ✓ Net foam accumulation



Key issues

- ✓ Reactor design and operational conditions
- ✓ Surfactant
- ✓ Microalgae culture

BACKGROUND

What are the desirable conditions for producing algae-rich foam?

- ✓ Suitable choice of surfactant + microalga strain
- ✓ Microalgae hold-up → Algae-enriched foam
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- ✓ Stable foam
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Key issues

- ✓ Reactor design and operational conditions
- ✓ Surfactant
- ✓ Microalgae culture

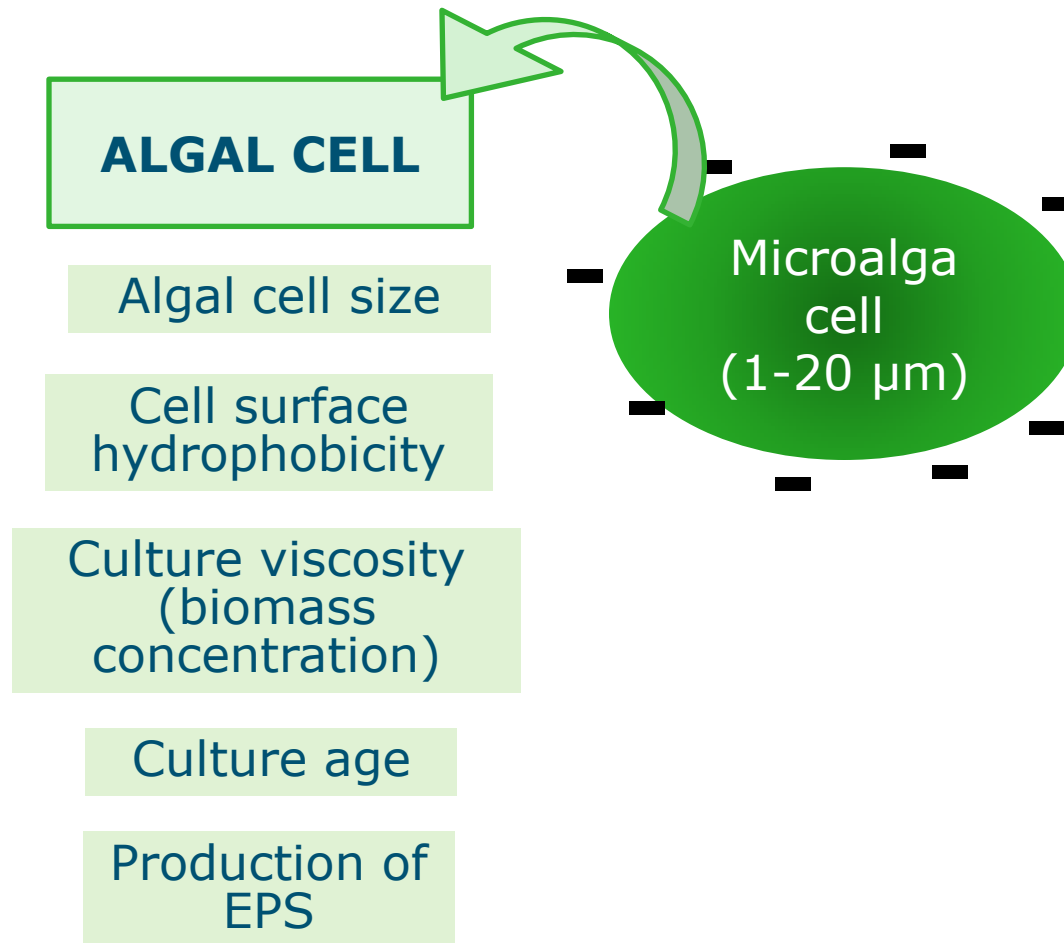


BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?

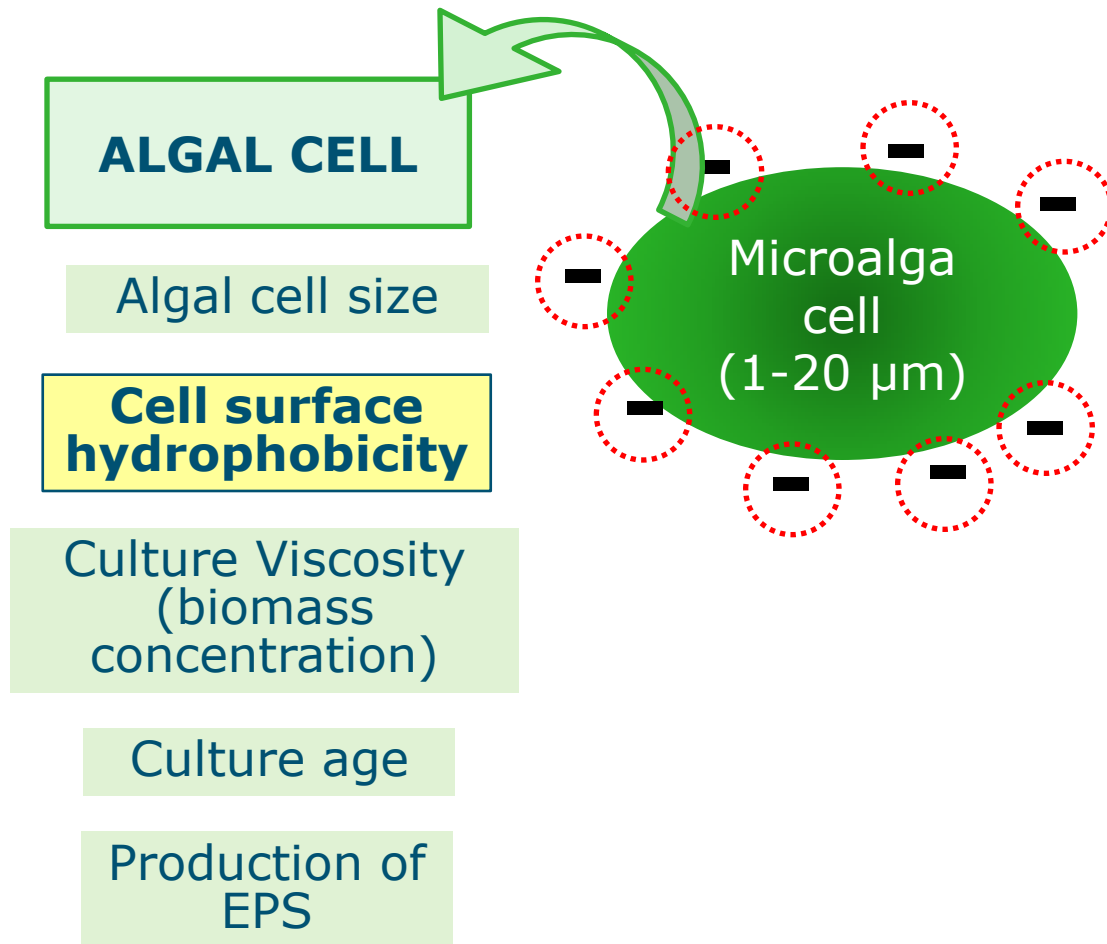
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What are the factors of the microalgae culture which influence the foam formation process?



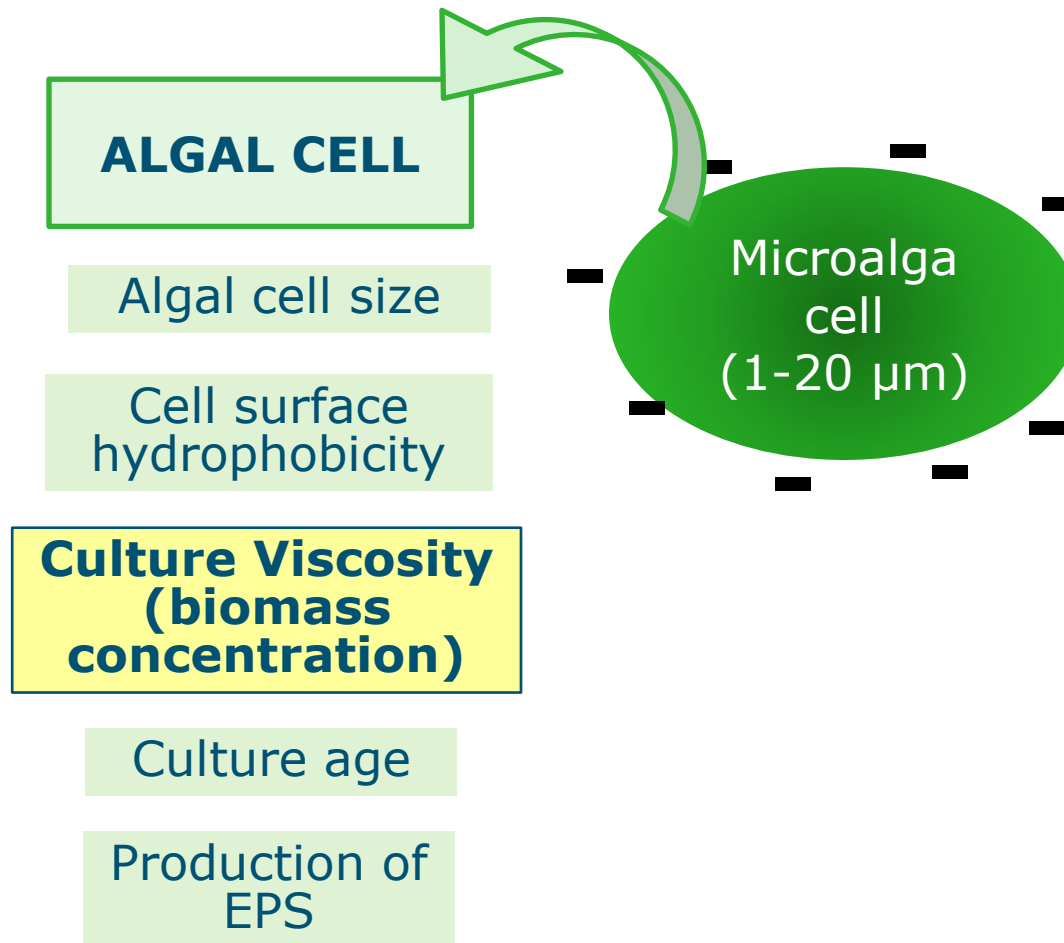
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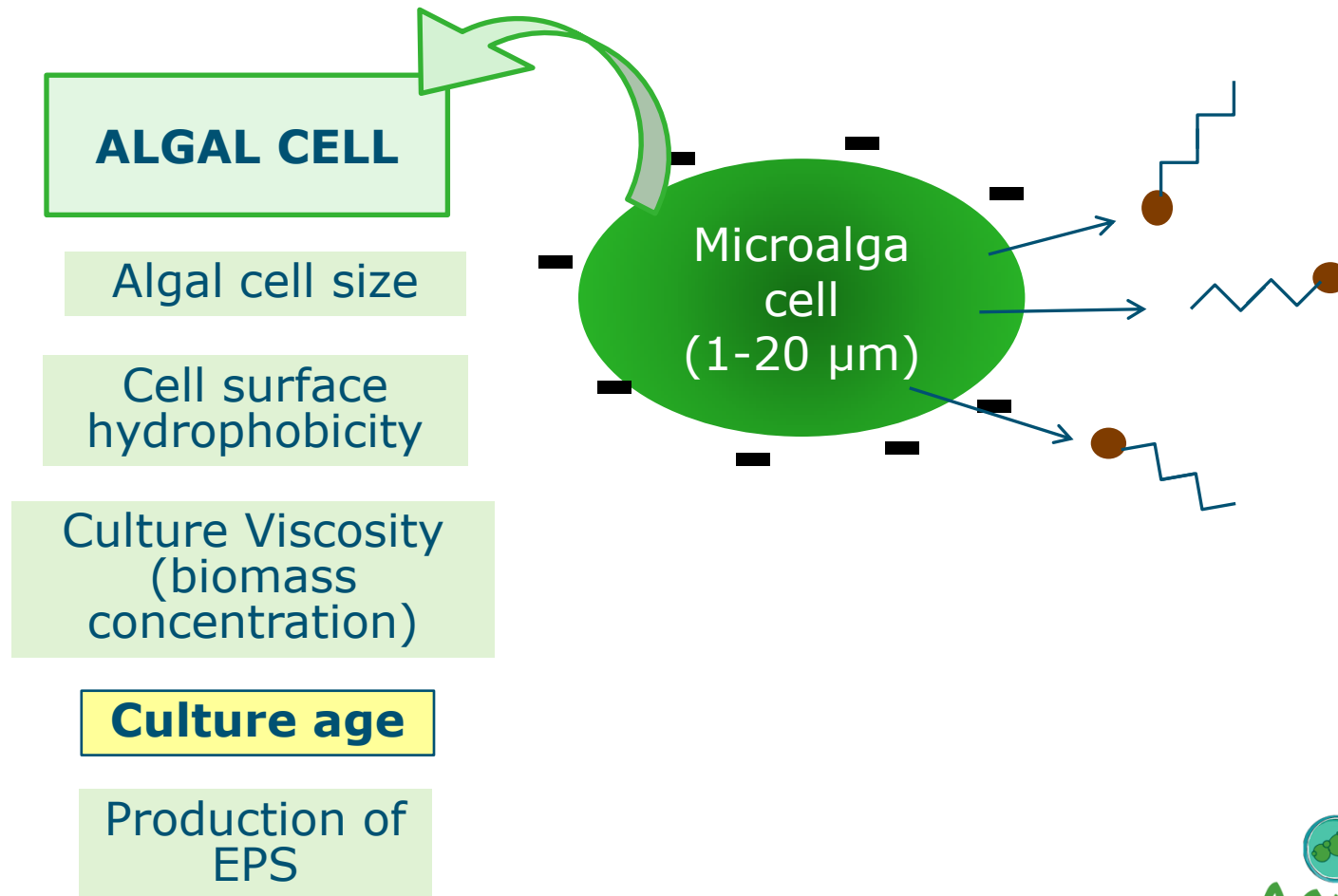
BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?



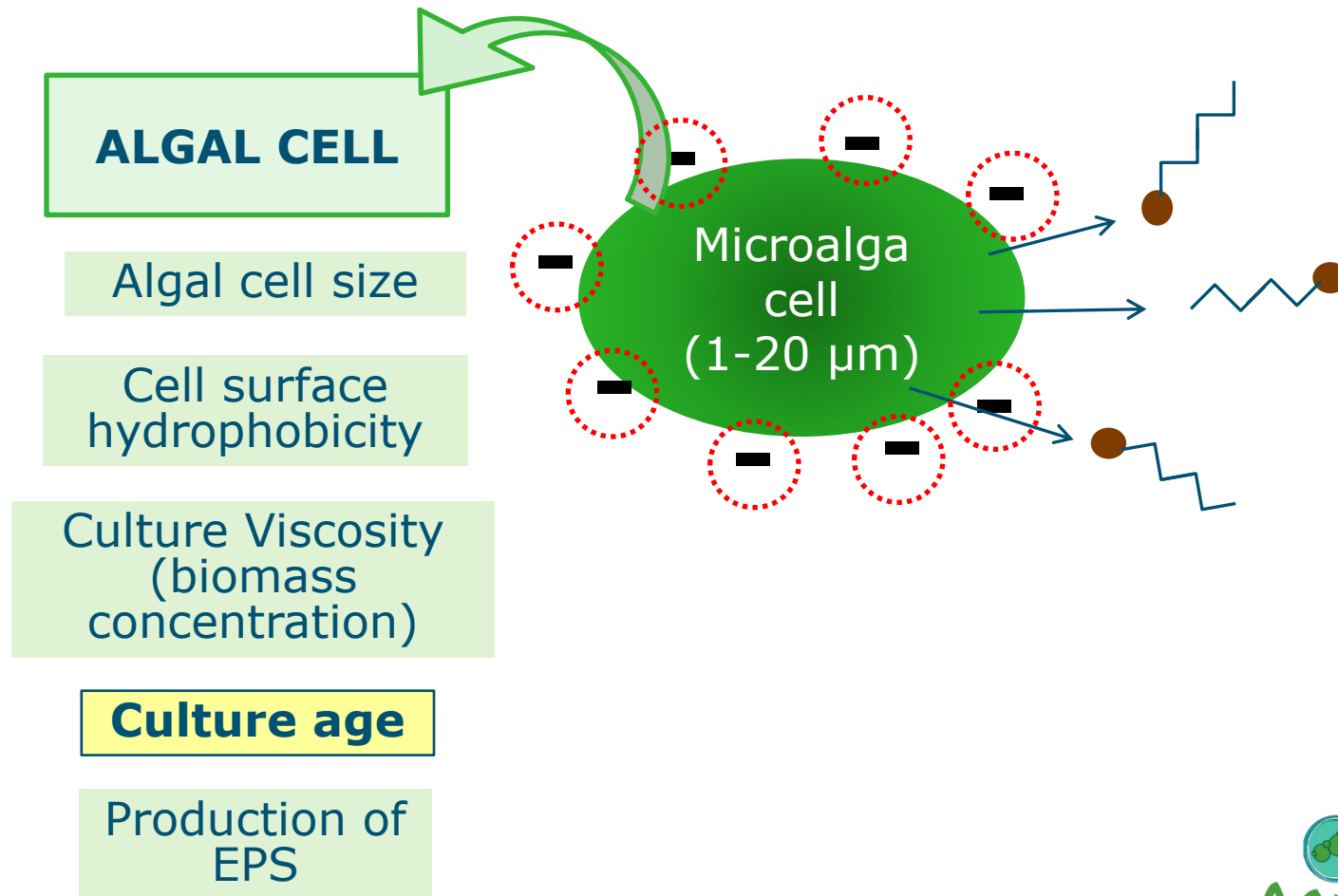
BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?



BACKGROUND

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BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?

ALGAL CELL

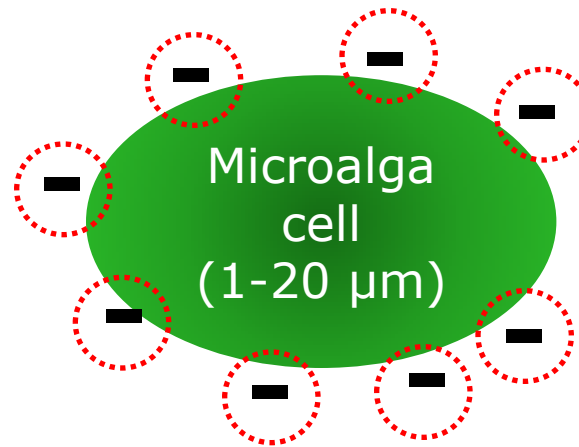
Algal cell size

Cell surface hydrophobicity

Culture Viscosity
(biomass concentration)

Culture age

Production of EPS



CULTURE MEDIUM

pH

Culture Medium salinity

BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?

ALGAL CELL

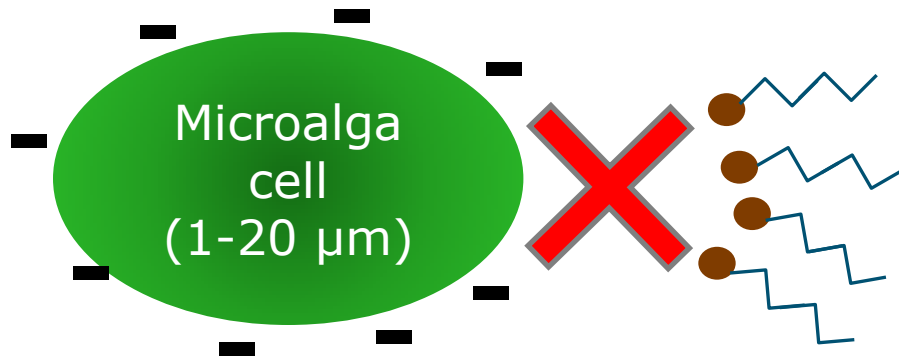
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Production of EPS



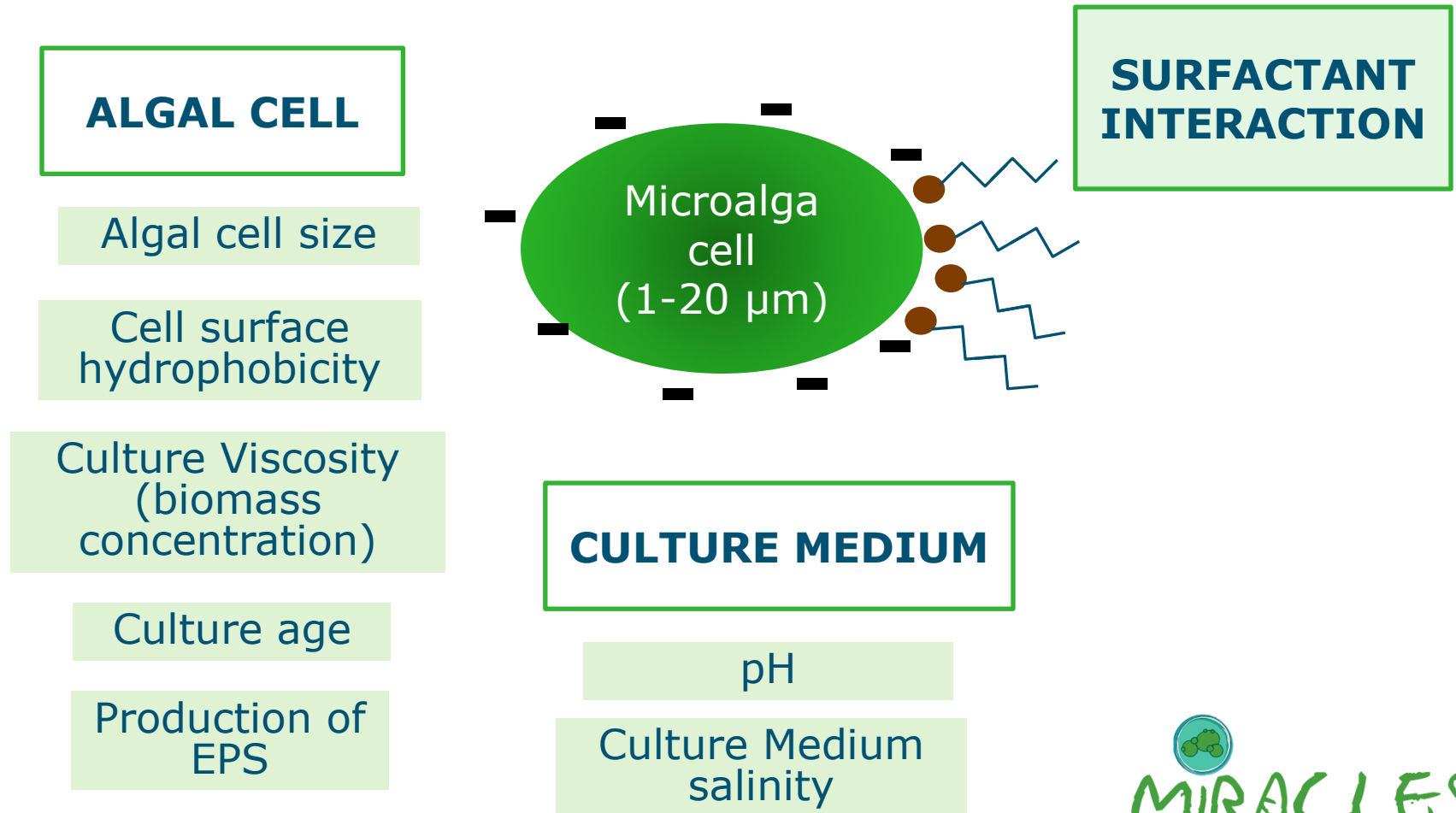
CULTURE MEDIUM

pH

**Culture Medium
salinity**

BACKGROUND

What are the factors of the microalgae culture which influence the foam formation process?



RESULTS

What have we done?

1. Preselection and production of **microalgae species** suitable for natural foam production.
2. Assessment of **natural foam production**
3. Comparison of surfactant-mediated **foam production speeds** (up-flow)
4. Study of the **hold-up ability**
5. Assessment of **foam stability**.

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RESULTS

1. Preselection and production of microalgae species suitable for natural foam production.

- 8 microalgae species
- Literature survey
- Strains included in the project
- Own experience and foaming observations

Microalga	Culture medium	Extracellular polymers	References
<i>Nannochloropsis gaditana</i> (CCFM)	F/2	Polysaccharides	Guerrero et al. 2004
<i>Scenedesmus obliquus</i> (CCFM)	BG-11	Proteins, polysaccharides	Corradi et al. 1998
<i>Phaeodactylum tricornutum</i> (CCFM)	F/2	Proteins, polysaccharides	Willis et al. 2013; Bruckner et al. 2011
<i>Isochrysis galbana</i> (CCFM)	F/2	-	-
<i>Chlorella sorokiniana</i>	M-8a	Proteins, polysaccharides	Wang et al. 2004; Campos et al. 2013
<i>Neochloris oleoabundans</i>	F/2	Biopolymers	Wu et al. 2011
<i>Botryococcus braunii</i> • var. Showa • (B14)	Chu	Hydrocarbons, exopolysaccharides	Dayananda et al. 2007

RESULTS

1. Preselection and production of microalgae species suitable for natural foam production.

Stable repeated-batch
cultures

Culture conditions

Light intensity:	160 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$
Temperature:	26 celsius
Mode:	Repeated-batch, log phase
Bubbling:	CO ₂ in air (5% v/v)
Volume:	2 to 5 L
Dense cultures:	Centrifugation



RESULTS

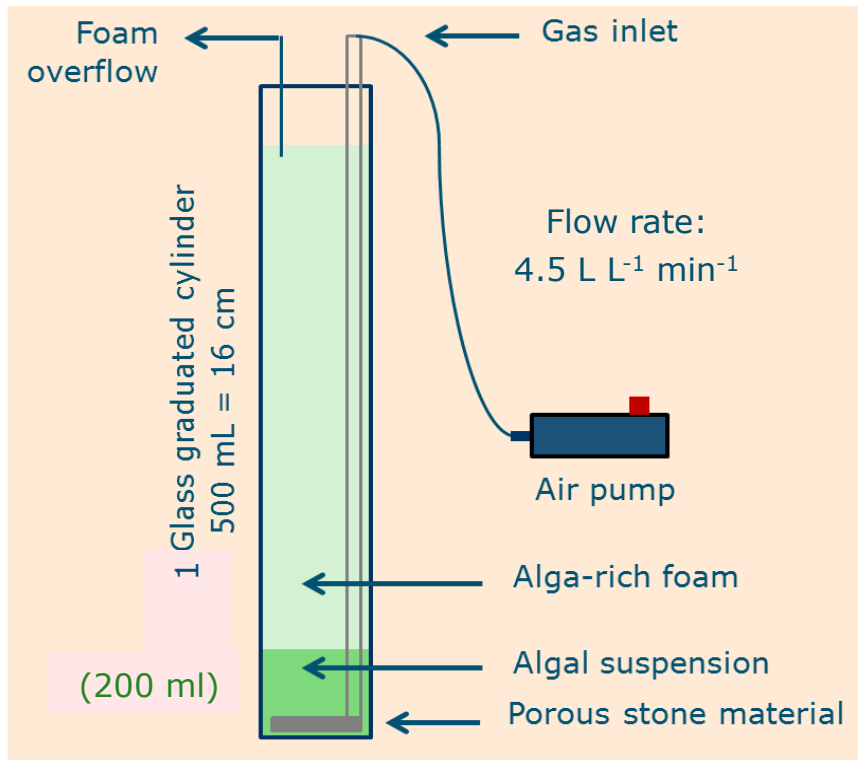
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RESULTS

2. Assessment of natural foam production

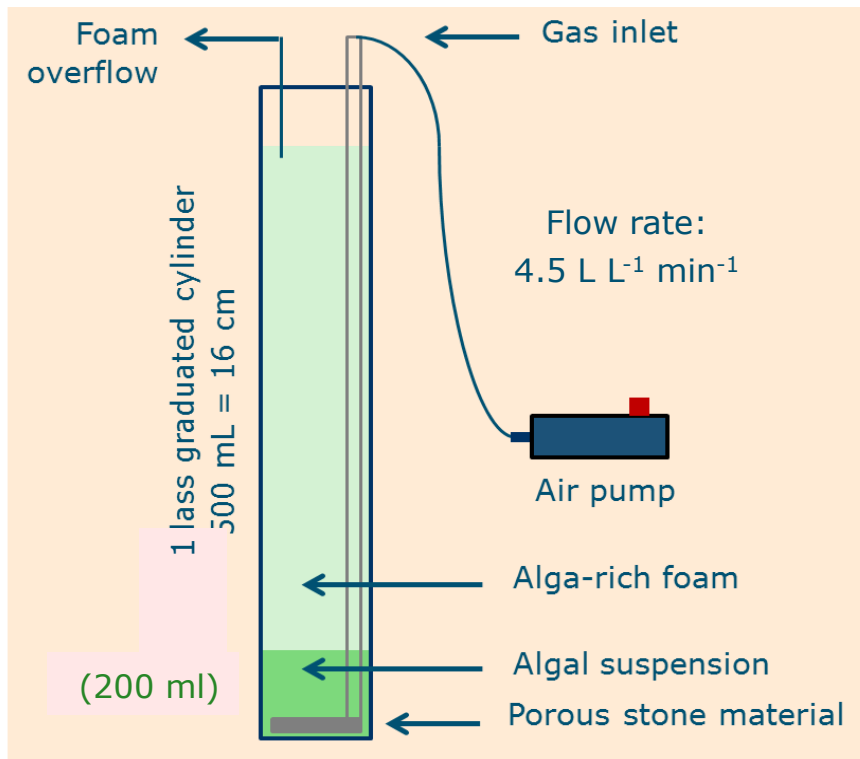
Comparing ability to produce foam naturally and in a fast way



RESULTS

2. Assessment of natural foam production

Comparing ability to produce foam naturally and in a fast way



Study parameters:

- Microalgae strains

- Freshwater

- *C. sorokiniana*
- *S. obliquus*,
- *B. braunii* var. Showa
- *B. braunii* (B14)

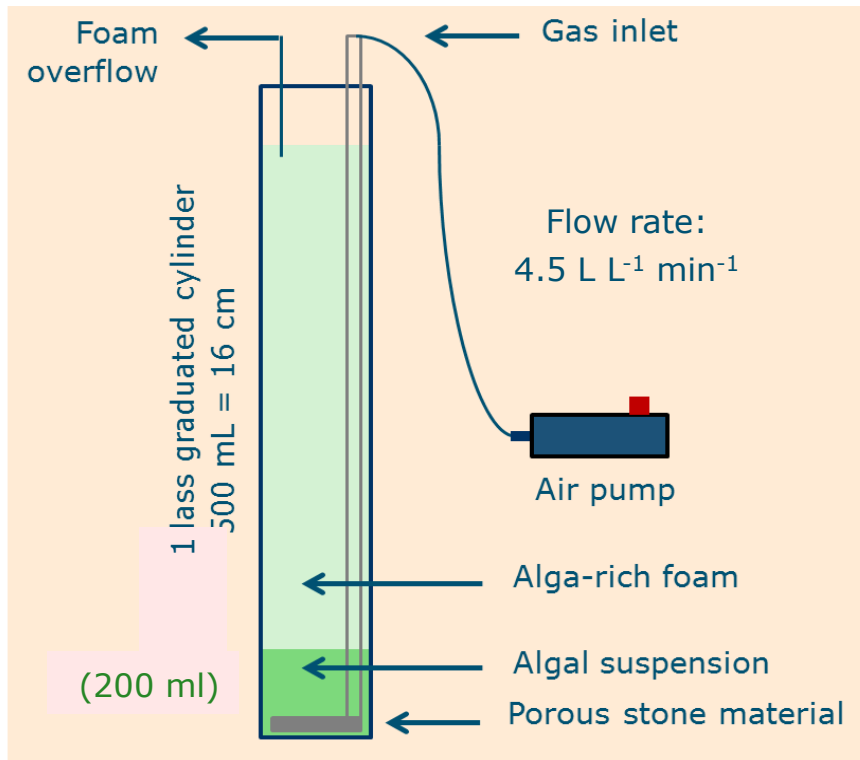
- Marine

- *I. galbana*
- *N. gaditana*
- *P. tricornutum*
- *N. oleoabundans*

RESULTS

2. Assessment of natural foam production

Comparing ability to produce foam naturally and in a fast way



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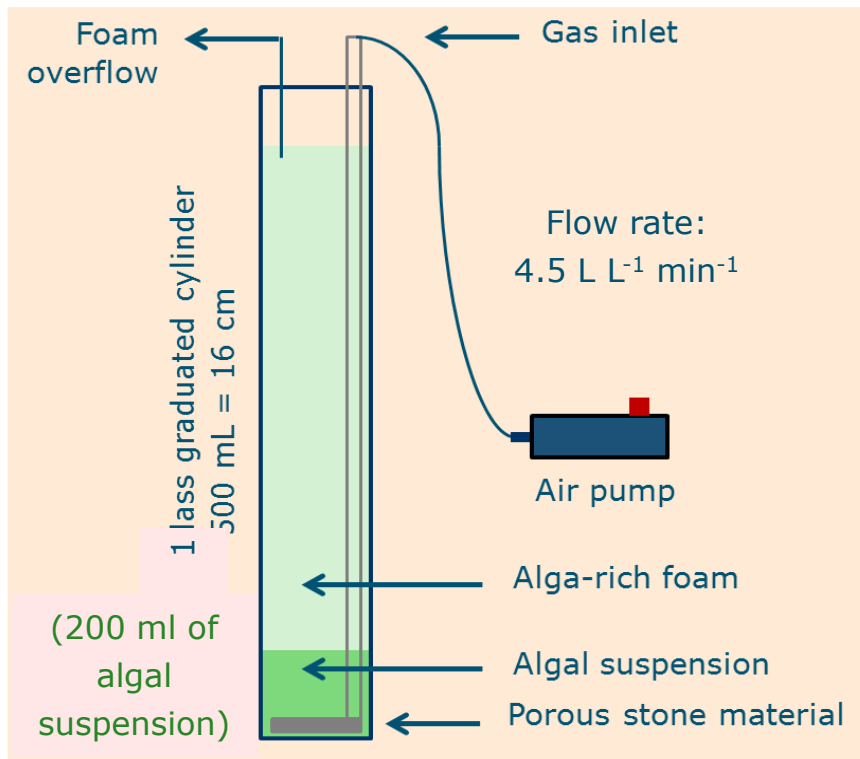
- Growth phase

- Exponential
- Stationary

RESULTS

2. Assessment of natural foam production

Comparing ability to produce foam naturally and in a fast way



Study parameters:

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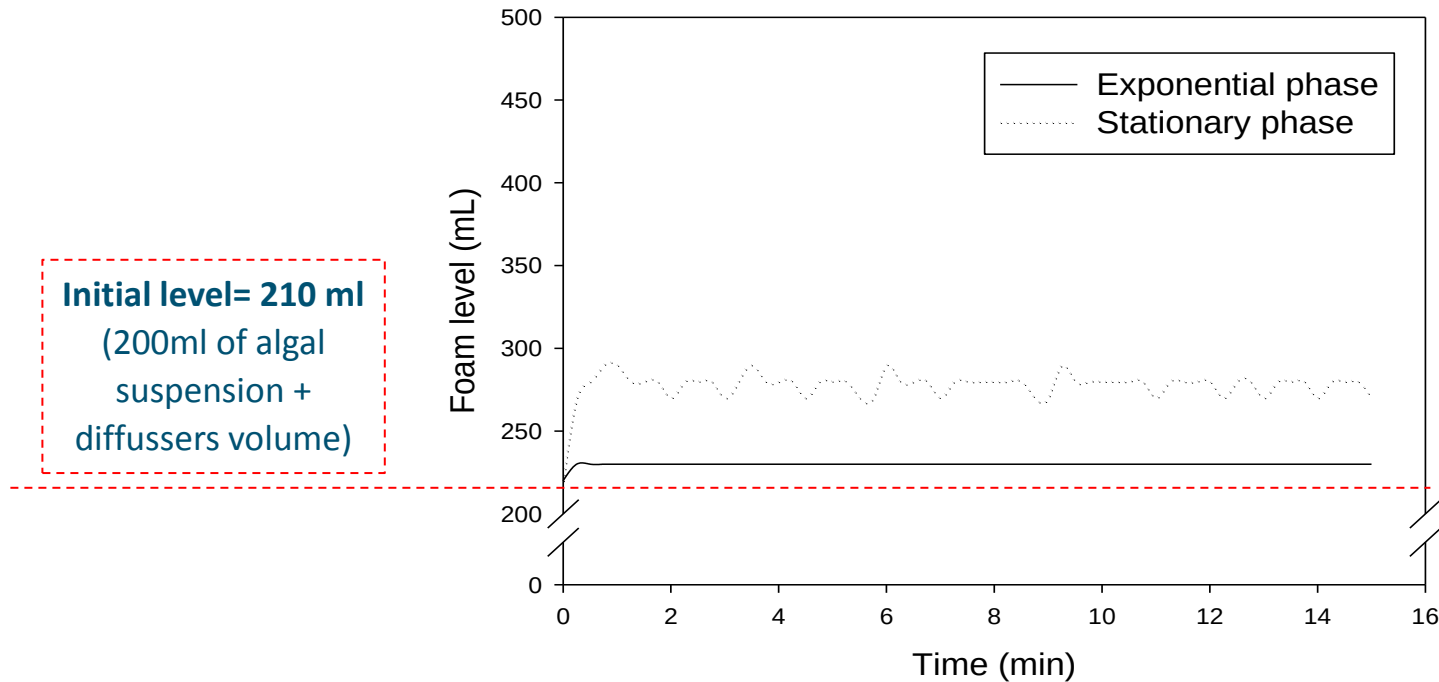
- Biomass concentration

- Normal
- 3-time concentrated

RESULTS

Effect of growth phase on natural foaming

N. oleoabundans, not concentrated

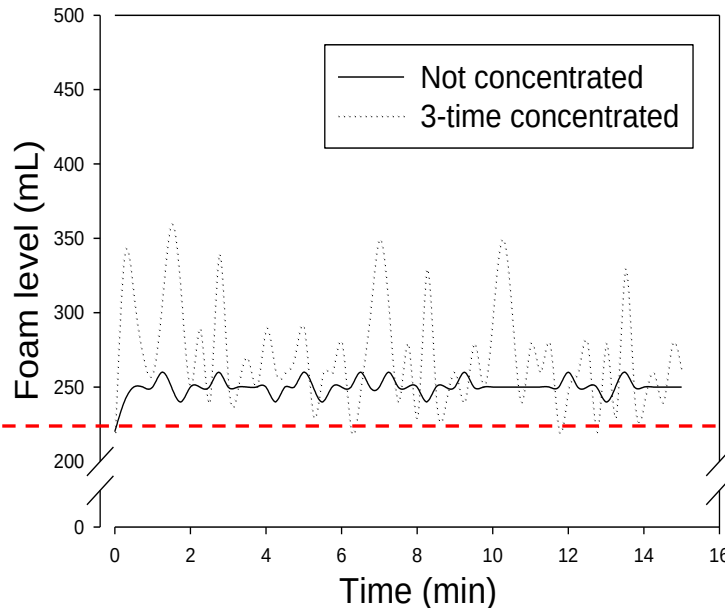


- ✓ **Stationary > Exponential:** *N. oleoabundans*, *B. braunii* var. Showa, *N. gaditana*
- ✓ **Foaming is preferably in exponential phase**

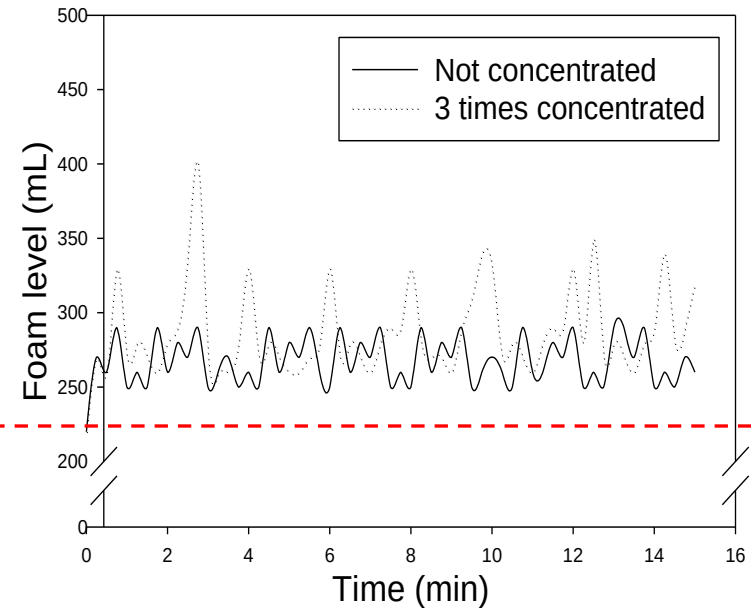
RESULTS

Effect of biomass concentration on natural foaming

B. braunii var. Showa (stationary phase)



B. braunii (B14) (stationary phase)

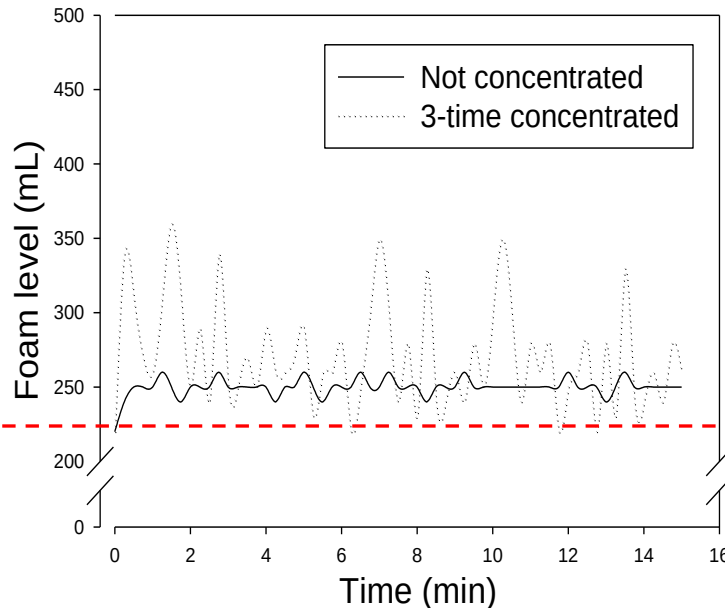


✓ **Positive effect:** *B. braunii* var. Showa and *B. braunii* (B14)

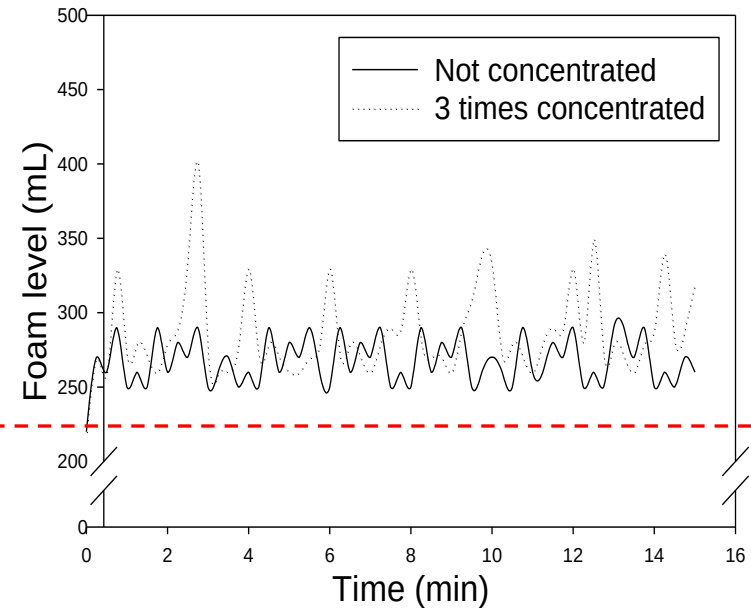
RESULTS

Effect of biomass concentration on natural foaming

B. braunii var. Showa (stationary phase)

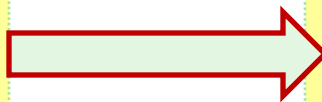


B. braunii (B14) (stationary phase)



✓ **Positive effect:** *B. braunii* var. Showa and *B. braunii* (B14)

**Not stable foam naturally
produced in any case.**



**SURFACTANT ADDITION IS
REQUIRED !!!**

RESULTS

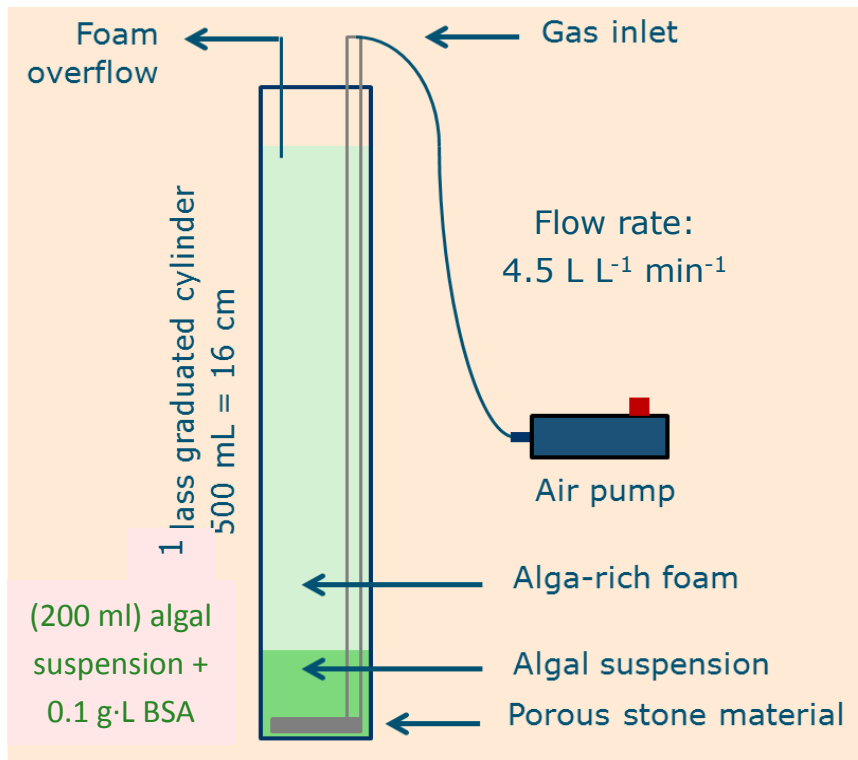
What have we done?

1. Preselection and production of **microalgae species** suitable for natural foam production.
2. Assessment of natural foam production
3. Comparison of **surfactant-mediated foam production speeds (up-flow)**
4. Study of the **up-hold ability**
5. Assessment of the **foam stability**.

RESULTS

3. Surfactant-mediated foam production speeds (up-flow)

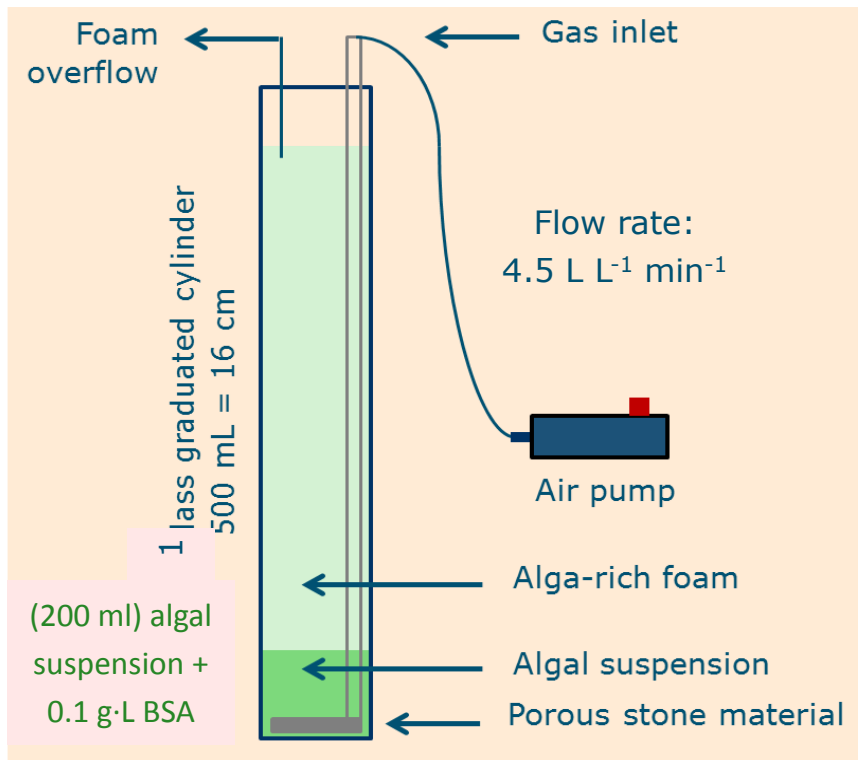
Comparing ability to produce BSA-mediated foaming in a fast way



RESULTS

3. Surfactant-mediated foam production speeds (up-flow)

Comparing ability to produce BSA-mediated foaming in a fast way



Study parameters:

- Microalgae strains

- Freshwater
 - *C. sorokiniana*
 - *S. obliquus*,
 - *B. braunii* var. Showa
 - *B. braunii* (B14)
- Marine
 - *I. galbana*
 - *N. gaditana*
 - *P. tricornutum*
 - *N. oleoabundans*

- Growth phase

- Exponential
- Stationary

- Biomass concentration

- Normal
- 3-time concentrated

RESULTS

3. Surfactant-mediated foam production speeds (up-flow)



BSA: Bovine Serum Albumin

- ✓ Organic
- ✓ Non-toxic
- ✓ Efficacy as surfactant previously shown

RESULTS

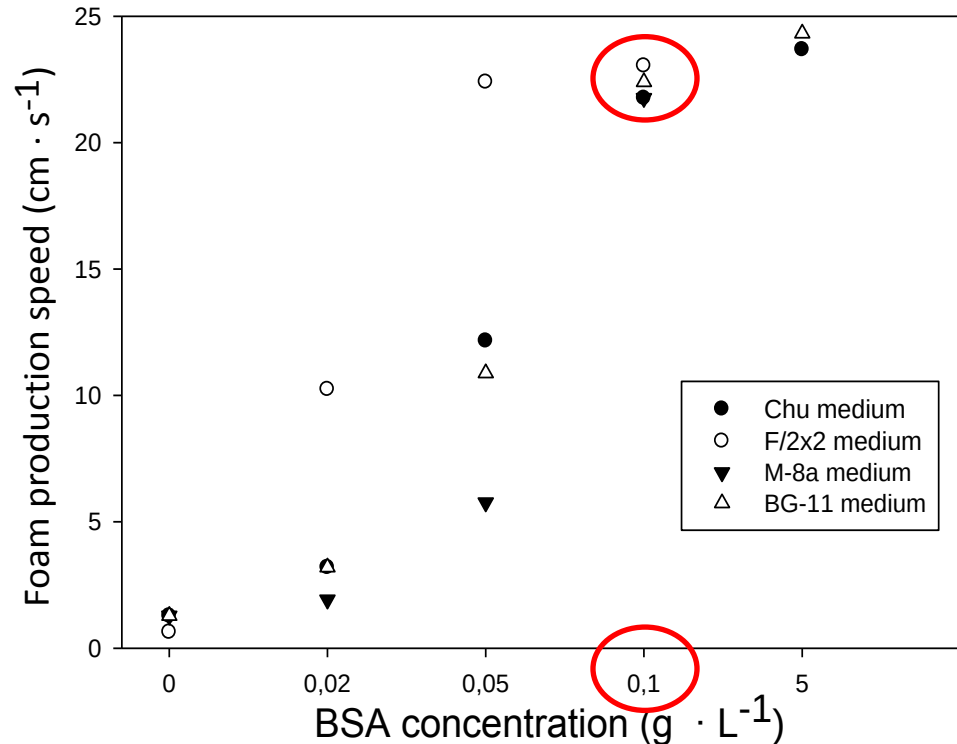
3. Surfactant-mediated foam production speeds (up-flow)



BSA: Bovine Serum Albumin

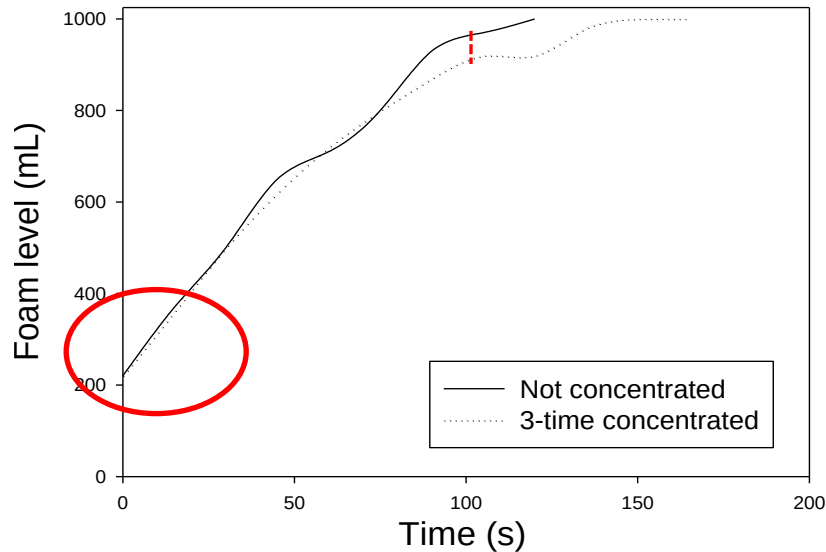
- ✓ Organic
- ✓ Non-toxic
- ✓ Efficacy as surfactant previously shown
- ✓ **$0.1 \text{ g} \cdot \text{L}^{-1}$**

BSA concentration assessment

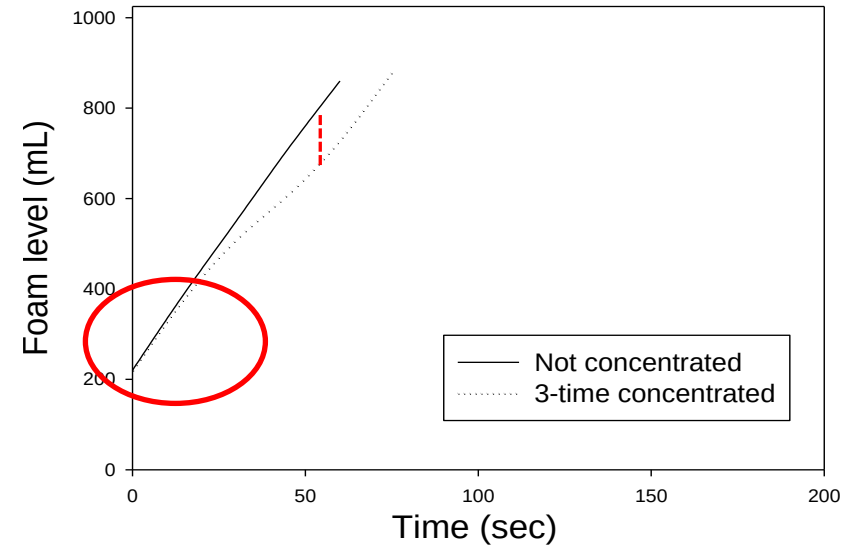


Effect of biomass concentration on BSA-mediated foam up-flow

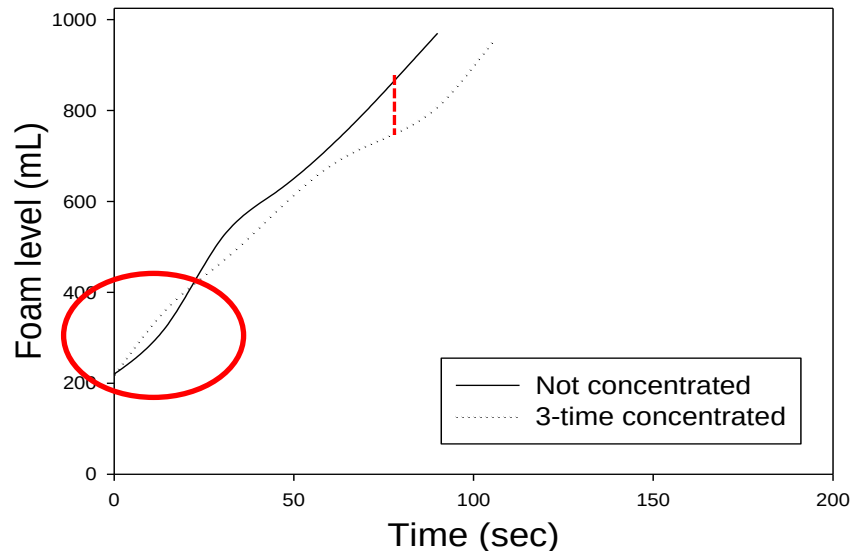
B. braunii (B14), stationary phase



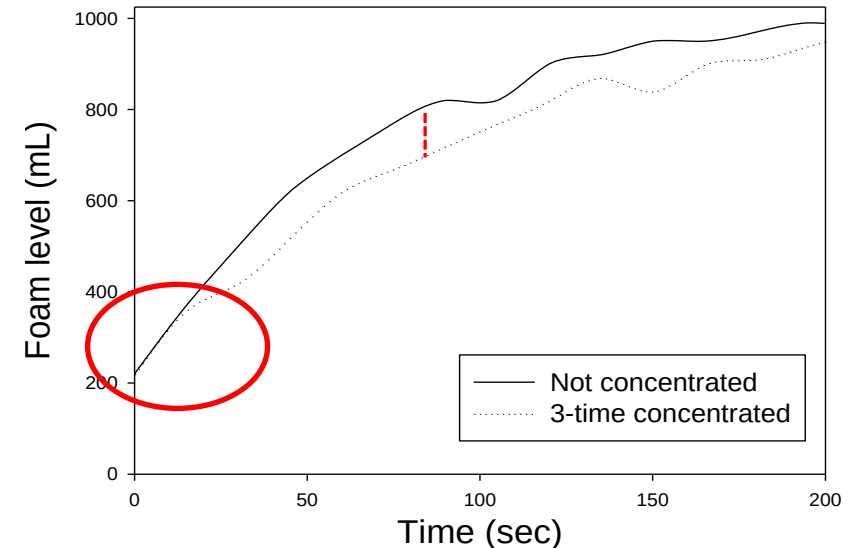
C. sorokiniana, exponential phase



N. oleoabundans, stationary phase



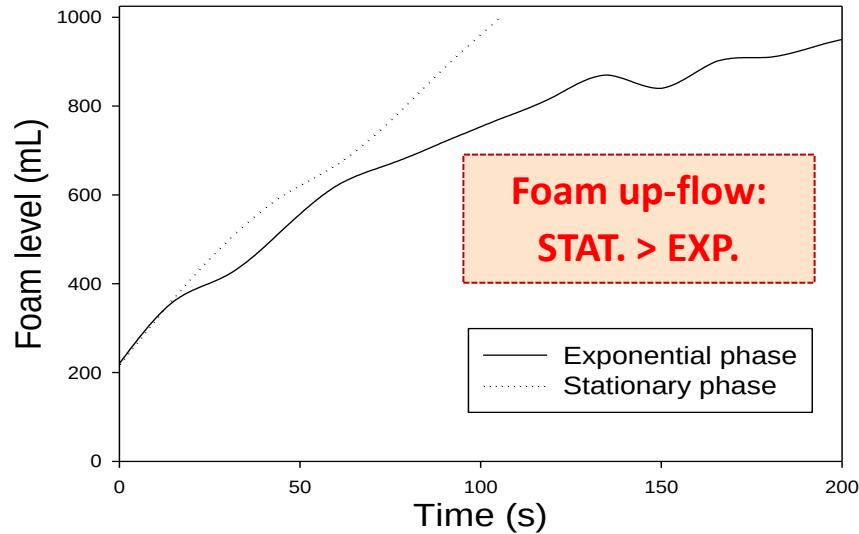
N. gaditana, exponential phase



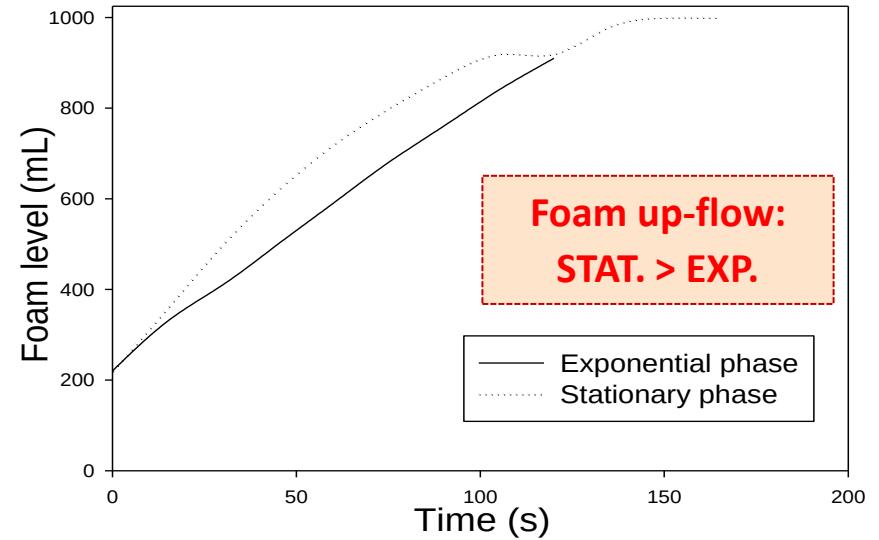
Biomass preconcentration led to lower up-flow just seconds after foaming started up

Effect of growth phase on BSA-mediated foam up-flow

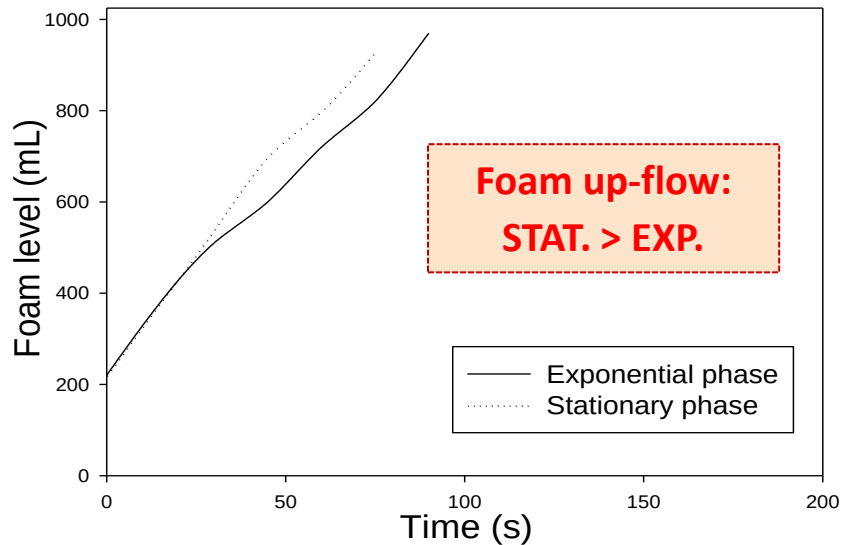
N. gaditana, 3-time concentrated



B. Braunii (B14), 3-time concentrated



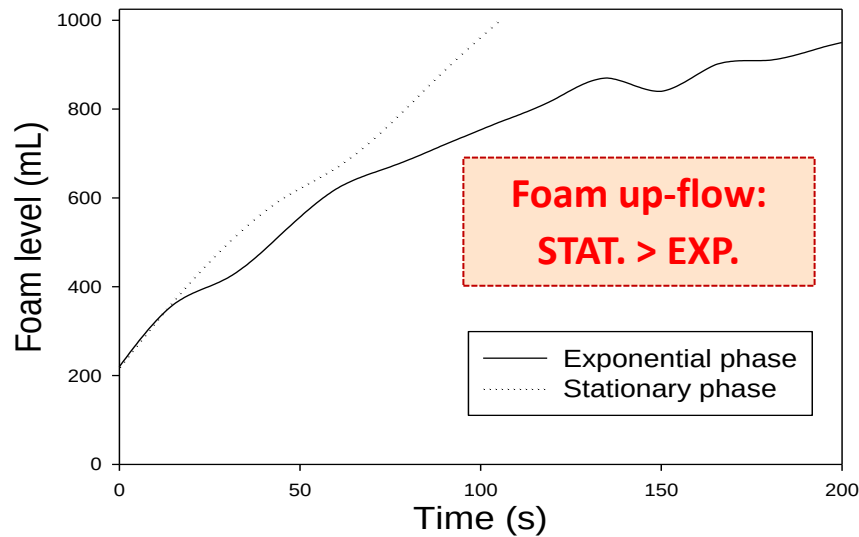
S. obliquus, 3-time concentrated



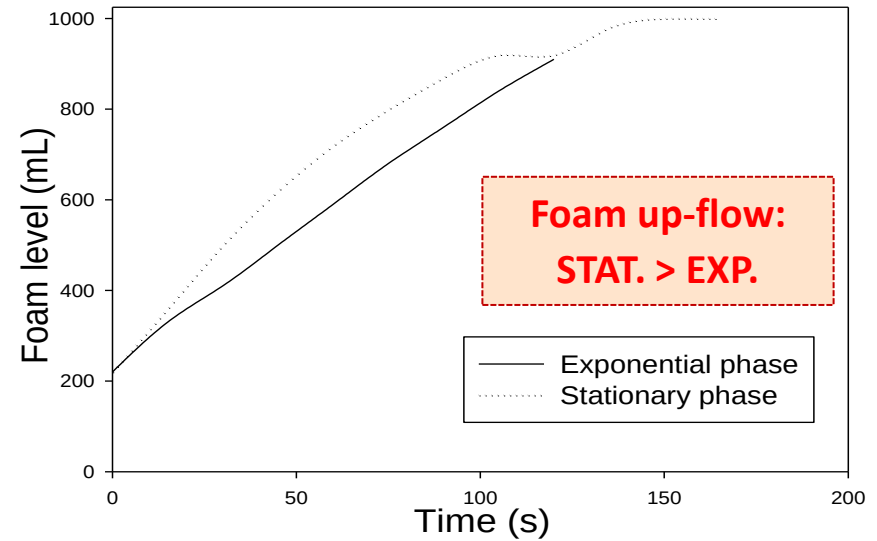
**Stationary phase
favoured foaming in
most cases**

Effect of growth phase on BSA-mediated foam up-flow

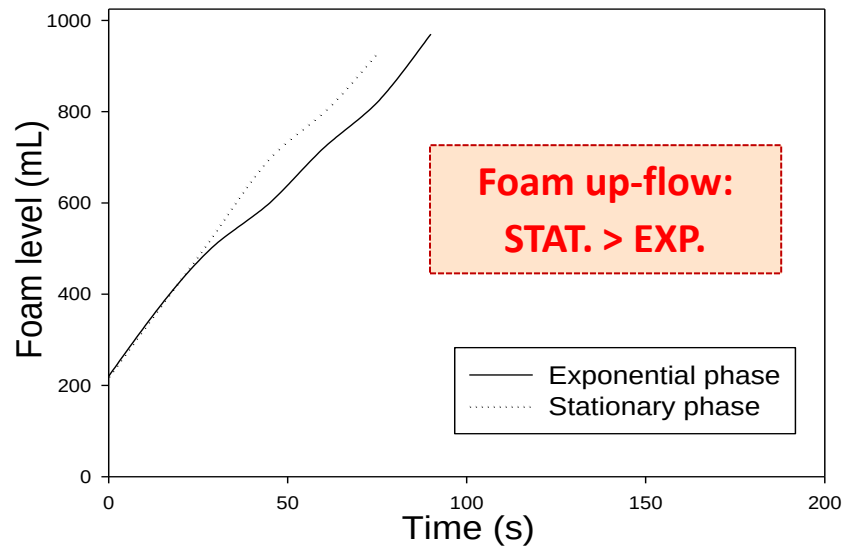
N. gaditana, 3-time concentrated



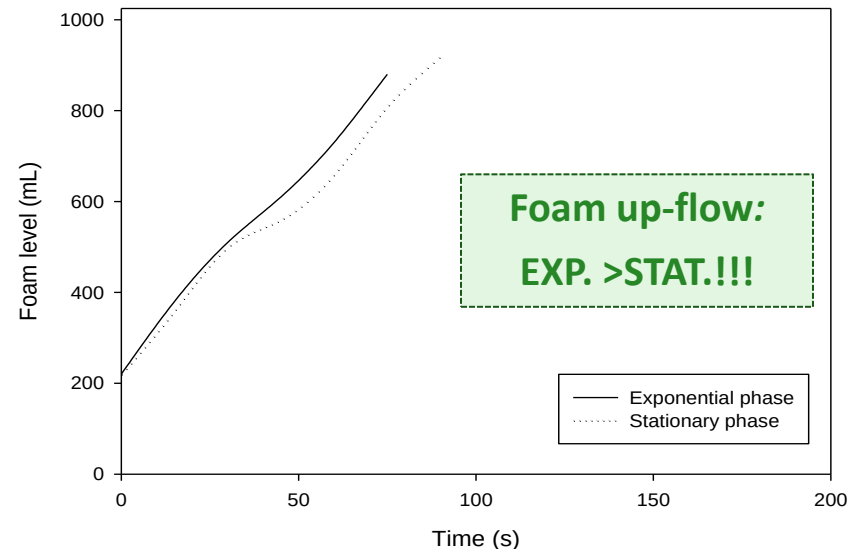
B. Braunii (B14), 3-time concentrated



S. obliquus, 3-time concentrated

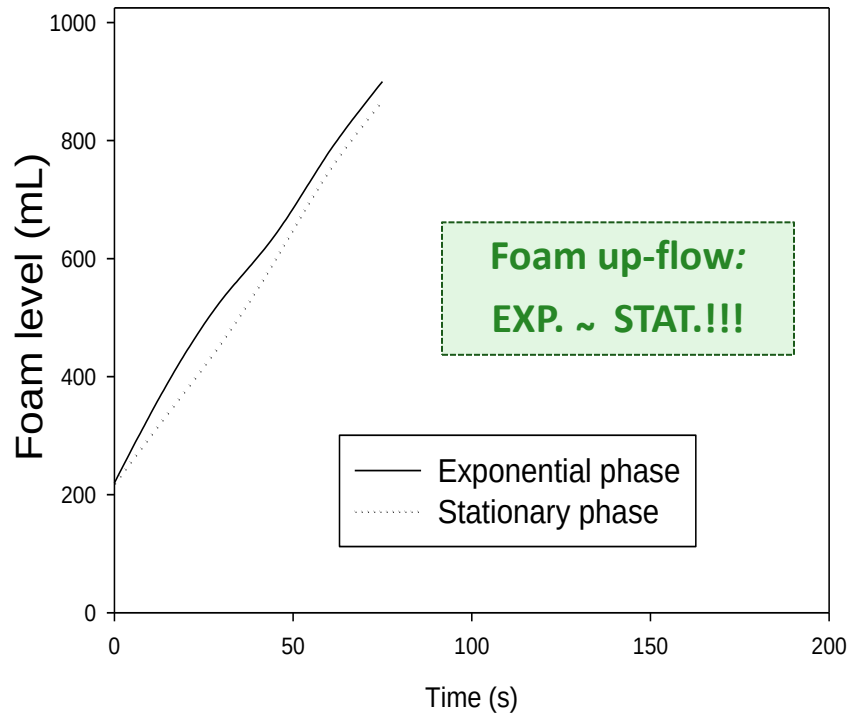


C. sorokiniana, 3-time concentrated



***B. braunii* var. *Showa* produces foam naturally, even in exponential phase...**

B. braunii var. *Showa*, not concentrated



However... *Showa* strain will not be considered as a suitable strain to produce algae-rich foam because of its low hold-up.

BSA-mediated foam up-flow speeds (cm · s⁻¹)

	Chu medium	F/2x2 medium	<i>Chlorella sorokiniana</i>		<i>Botryococcus braunii</i> var. Showa	
	NO ALGAE	NO ALGAE	LOW ALGAE CONC.	HIGH ALGAE CONC.	LOW ALGAE CONC.	HIGH ALGAE CONC.
Exponential	0.36	0.38	0,35	0.31	0,33	0,32
Stationary			0,32	0.30	0,26	0.26
	<i>Botryococcus braunii</i> (B14)		<i>Scenedesmus obliquus</i>		<i>Nannochloropsis gaditana</i>	
	LOW ALGAE CONC.	HIGH ALGAE CONC.	LOW ALGAE CONC.	HIGH ALGAE CONC.	LOW ALGAE CONC.	HIGH ALGAE CONC.
Exponential	0,21	0,20	0,31	0,31	0,30	0,21
Stationary	0,30	0,30	0,33	0,34	0,30	0,30
	<i>Isochrysis galbana</i>		<i>Neochloris oleoabundans</i>		<i>Phaeodactylum tricornutum</i>	
	LOW ALGAE CONC.	HIGH ALGAE CONC.	LOW ALGAE CONC.	HIGH ALGAE CONC.	LOW ALGAE CONC.	HIGH ALGAE CONC.
Exponential	N.M.	N.M.	0,23	0,23	0,27	0,32
Stationary	0,18	0,13	0,32	0,27	N.M.	N.M.

RESULTS

What have we done?

1. Preselection and production of **microalgae species** suitable for natural foam production.
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RESULTS

4. Study of algae hold-up abilities

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RESULTS

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- Growth phase

- Exponential
- ~~Stationary~~

RESULTS

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- *P. tricornutum*
- *N. oleoabundans*

- Growth phase

- Exponential
- Stationary

- Biomass concentration

- LOW BIOMASS: $1.5-2 \text{ g} \cdot \text{L}^{-1}$
- HIGH BIOMASS: $8 - 10 \text{ g} \cdot \text{L}^{-1}$

RESULTS

4. Study of algae hold-up abilities

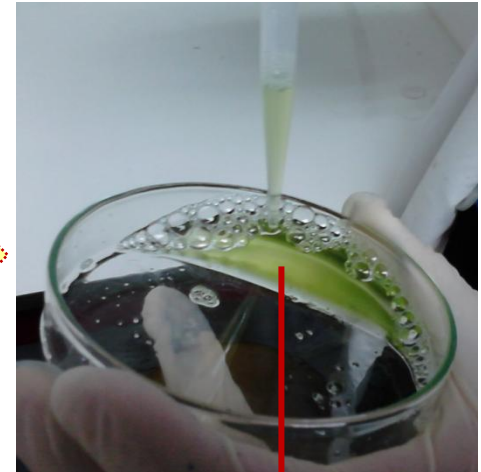
Comparing hold-up abilities



D.W. (g·L⁻¹)_{liquid suspension}



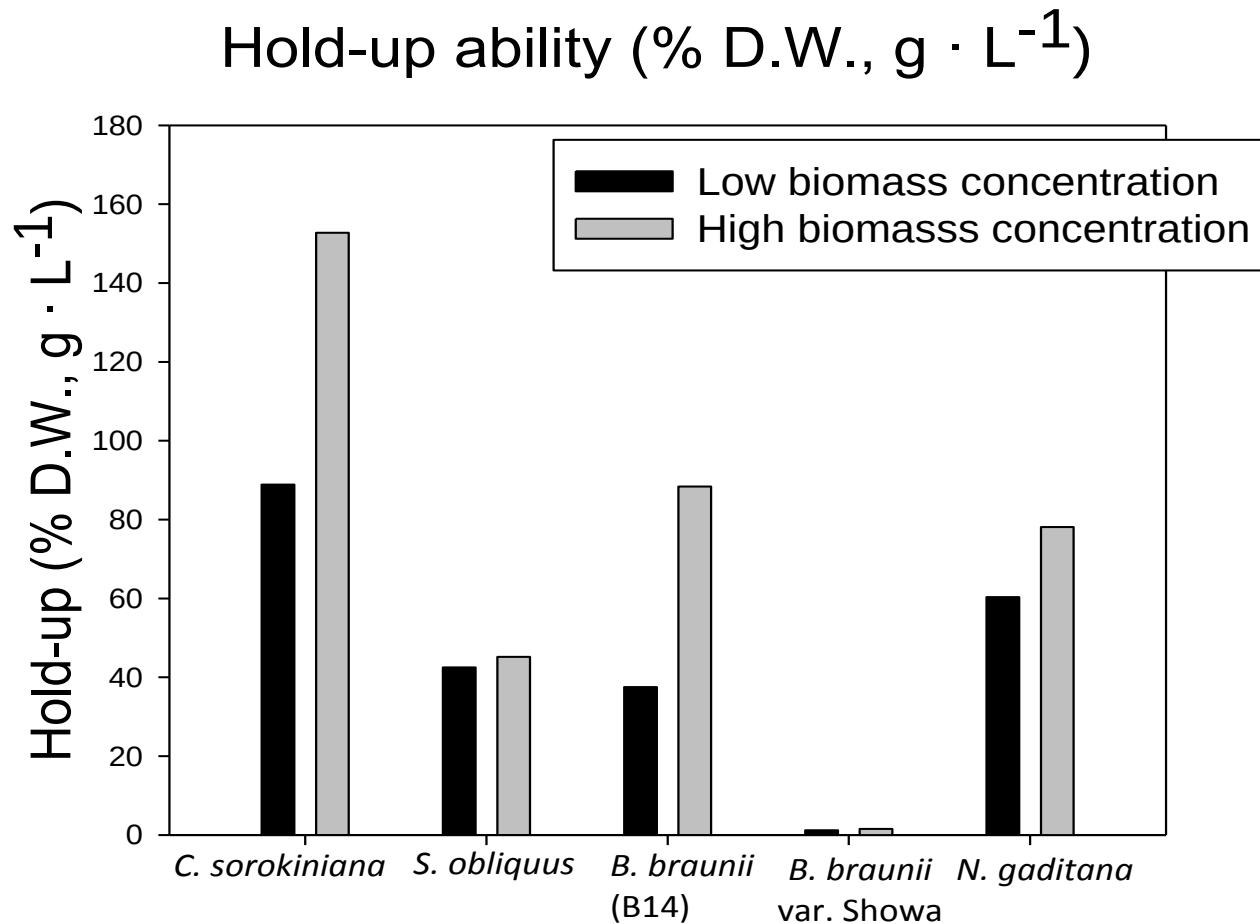
D.W. (g·L⁻¹)_{liquid from the foam}



$$\text{Hold-up (\%)} = \frac{\text{D.W. (g·L}^{-1}\text{)}_{\text{liquid from the foam}}}{\text{D.W. (g·L}^{-1}\text{)}_{\text{liquid suspension}}} \times 100$$

RESULTS

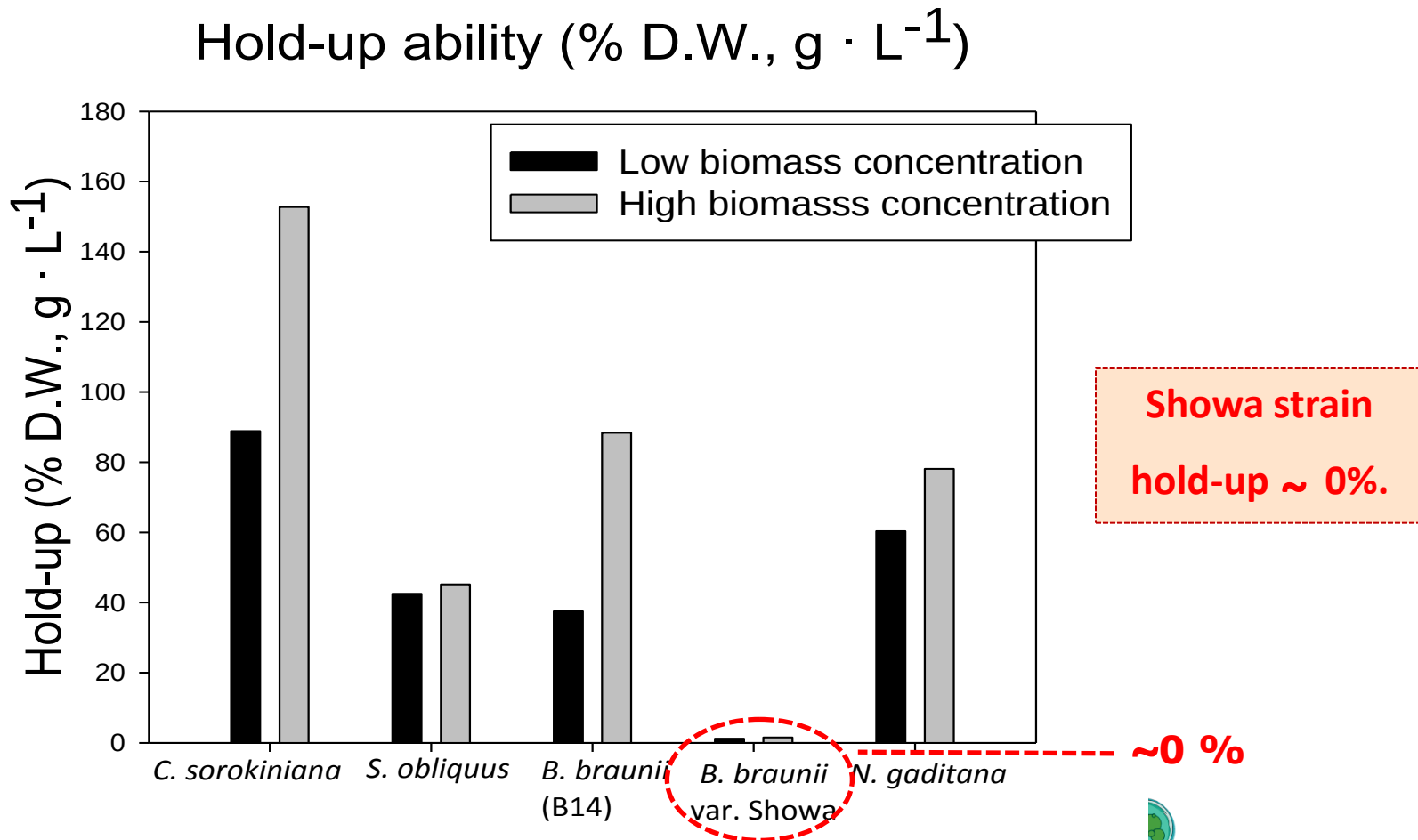
4. Study of algae hold-up abilities



Biomass preconcentration always favoured algae hold-up abilities.

RESULTS

4. Study of algae hold-up abilities



Biomass preconcentration always favoured algae hold-up abilities.

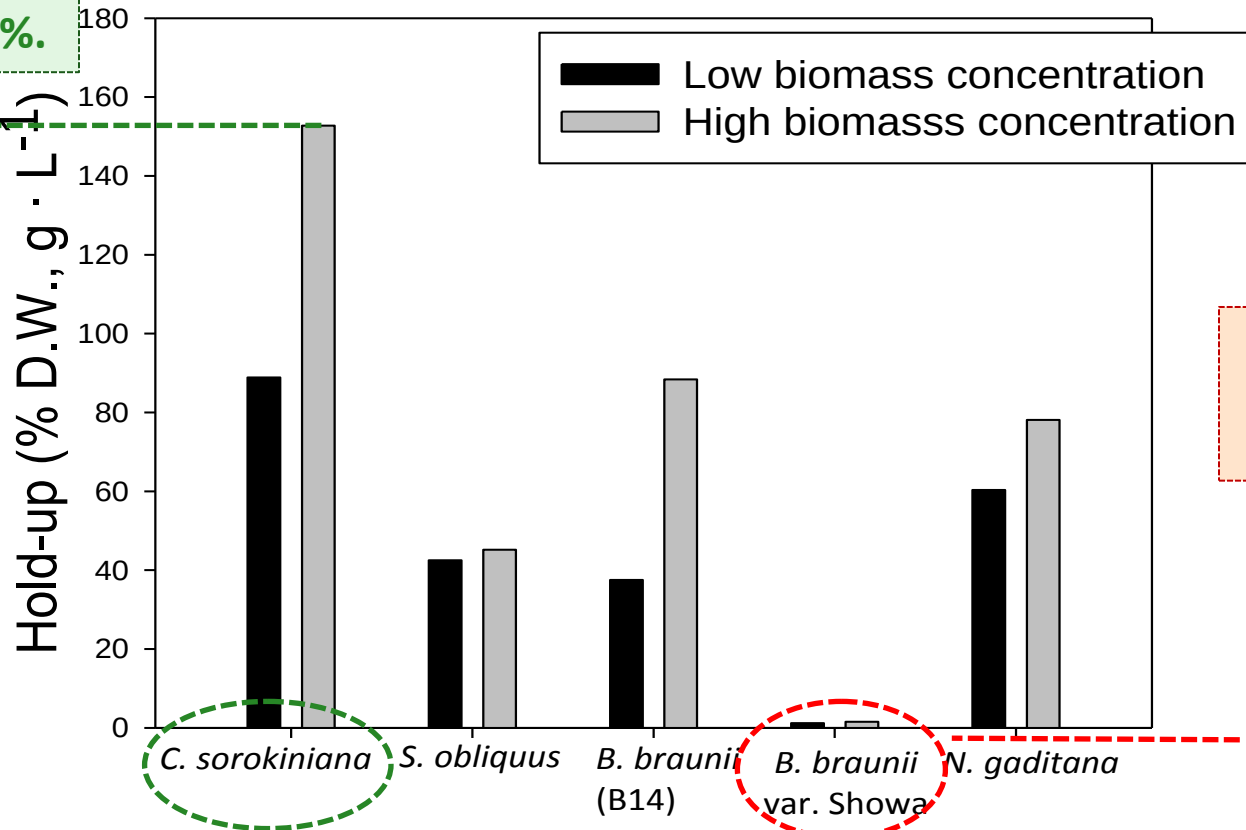
RESULTS

4. Study of algae hold-up abilities

Hold-up ability (% D.W., $\text{g} \cdot \text{L}^{-1}$)

Chlorella
hold-up > 100%.

~150 %



Showa strain
hold-up ~ 0%.

~0 %

Biomass preconcentration always favoured algae hold-up abilities.

RESULTS

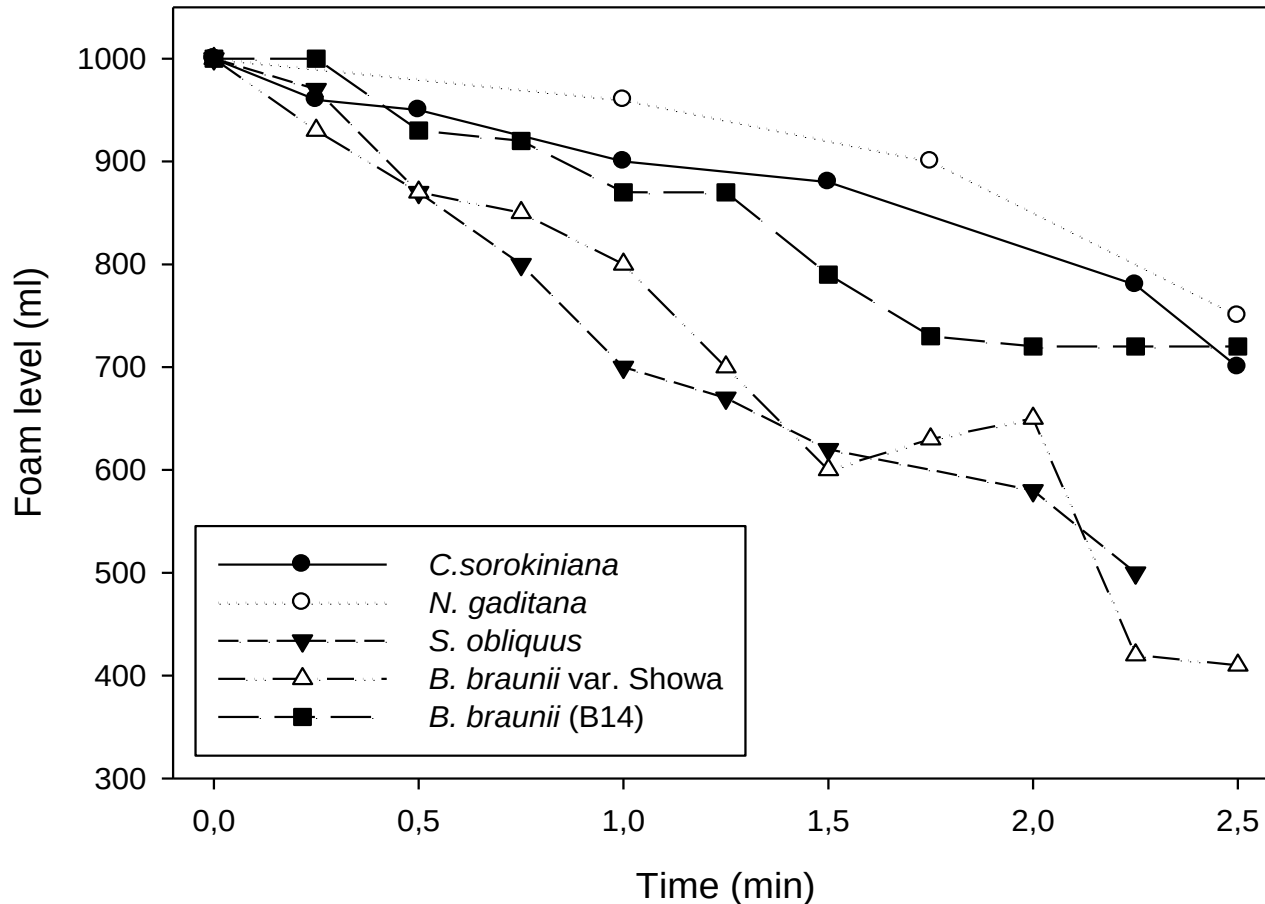
What have we done?

1. Preselection and production of **microalgae species** suitable for natural foam production.
2. Assessment of natural foam production
3. Comparison of surfactant-mediated foam production speeds (up-flow)
4. Study of hold-up ability
5. Assessment of **foam stability**.

RESULTS

5. Assessment of foam stability

Foam desestabilisation



BEST RESULTS:

- *C. sorokiniana*
- *N. gaditana*
- *B. braunii* (B14)

NOT MEASURABLE:

- *N. oleoabundans*
- *I. galbana*

CONCLUSIONS

- For all microalgae tested, the low stability of naturally produced foam prevents its accumulation. Therefore, addition of a surfactant is required to produce microalgae-enriched foam.
- Some microalgae show higher foaming abilities when collected from stationary phase. However, old cultures are not desirable for continuous cultivation.
- Due to low foam stability or low hold-up, *Neochloris oleabundans*, *Isochrysis galbana* and *Botryococcus braunii* var. Showa will not be considered for future experiments.
- For exponential microalgal cultures, the most promising results of foam up-flow, microalgae hold-up and foam stability were obtained for *C. sorokiniana*, *N. gaditana*, *S. obliquus* and *B. braunii* (B14).
- These species will be employed in further studies.

ACKNOWLEDGEMENTS

**THANK YOU FOR
YOUR ATTENTION**



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